

UNIVERSITI PUTRA MALAYSIA

ORGANOGENESIS, SOMATIC EMBRYOGENESIS AND REGENERATION OF IMMATURE MALE FLOWERS OF BANANA CULTIVARS

KEYNOOSH KASHEFI

FP 2008 6



ORGANOGENESIS, SOMATIC EMBRYOGENESIS AND REGENERATION OF IMMATURE MALE FLOWERS OF BANANA CULTIVARS

By

KEYNOOSH KASHEFI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Agricultural Science

May 2008



Dedicated to:

My kind parents, whom I am indebted with all love My beloved spouse My sweet daughter: Nikta



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Agricultural Science

ORGANOGENESIS, SOMATIC EMBRYOGENESIS AND REGENERATION OF IMMATURE MALE FLOWERS OF BANANA CULTIVARS

By

KEYNOOSH KASHEFI

May 2008

Chairman: Associate Professor Maheran Abdul Aziz, PhD

Faculty: Agriculture

This study was carried out to establish a reliable and practical protocol of plant regeneration through organogenesis and somatic embryogenesis of immature male flowers of four cultivars of *Musa* spp. cvs. Berangan, Rastali, Mas and Raja as well as micromorphological studies and optimization of plant transformation protocol via microprojectile bombardment of immature male flowers of *Musa* spp. cv. Rastali.

Immature male flowers are highly proliferated meristems which are ideal materials for organogenesis and embryogenesis in *Musa* spp. Immature male flowers of four cultivars of *Musa* spp. cvs. Berangan, Rastali, Mas and Raja were cultured on a modified MS medium containing 0, 9, 18 and 36 μ M BAP. The study showed a reasonably high percentage of adventitious bud induction on MS medium supplemented with 9 μ M BAP for cultivars Rastali and Raja and 18 μ M BAP for cultivars Berangan and Mas respectively within 4 to 8 weeks from the initiation of culture. Regeneration of shoots from the adventitious buds derived from immature male flowers of cultivars Berangan, Rastali, Mas and Raja were investigated on MS medium supplemented with 4.5, 9, 18



and 36 μ M BAP after four weeks of culture with a weekly subculture interval. It was observed that BAP at 4.5 µM produced the highest number of shoots and BAP at 36 µM produced the highest shoot-length for all cultivars tested after four subcultures. Subculturing significantly affected the mean number of shoots produced with the highest mean number of 40.33, 5.66, 23.66 and 19.00 shoots produced in cvs. Rastali, Raja, Berangan and Mas respectively in the third subculture (out of four subcultures). The mean shoot height also increased over subculture cycles in all cultivars.

Different coconut water preparations (filter-sterilized and autoclaved) in combination with the best BAP concentration for adventitious bud induction determined earlier for each cultivar were investigated. Highest mean number of adventitious buds after three subcultures (17.30) was attained on medium containing 50 mlL⁻¹ filter-sterilized coconut water combined with BAP at 9 µM for cvs. Rastali and Raja while 18 µM for cvs. Berangan and Mas.

Shoots obtained from the immature male flowers of cvs. Rastali, Raja, Berangan and Mas were rooted on half-strength MS medium supplemented with 1.0 μ M IBA and the plantlets produced were successfully acclimatized in the growth chamber.

Histological and Scanning Electron Microscopy (SEM) studies were carried out on male flowers of Musa spp. cv. Rastali placed on 9 µM BAP treatment at the initial stage of culture and the first, second and fourth subculture. Sequential changes were observed starting from globular mass like structures (bulges) to adventitious buds and finally producing multiple shoots.

Somatic embryogenesis of *Musa* spp. cv. Raja was established using immature male flower hands. Highest percentage of embryogenic callus formation (41.97%) was obtained on 13.5 μ M 2,4-D for all flower hand positions assessed. Flower hand position 8 produced the highest percentage (48.25%) of embryogenic callus formation for all levels of 2,4-D tested. The study revealed that the embryogenic cell suspensions initiated from the embryogenic callus/complex had high potential towards somatic embryogenesis. The highest percentage of somatic embryos germination (62.36%) was attained on medium with 0.17 μ M BAP after two weeks of culture.

Transformation study showed that the target distance of 9 cm along with helium pressure of 1350 and 1550 psi were the most efficient combinations for particle gun bombardment of immature male flower buds of cv. Rastali whereby 58.26% and 57.63% of the bombarded plates showed a high GFP gene expression.

Overall, this study indicated that immature male flower buds of *Musa* spp. cultivars Berangan, Rastali, Mas and Raja can be the appropriate materials for *in vitro* regeneration via organogenesis and somatic embryogenesis as well as for gene transformation via particle bombardment. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains Pertanian

ORGANOGENESIS, EMBRIOGENESIS SOMA DAN REGENERASI BUNGA JANTAN TIDAK MATANG BEBERAPA KULTIVAR PISANG

Oleh

KEYNOOSH KASHEFI

May 2008

Pengerusi: Profesor Madya Maheran Abdul Aziz, PhD

Fakulti: Pertanian

Kajian ini dilakukan untuk mendapatkan protokol yang sesuai dan praktikal bagi regenerasi menerusi organogenesis dan embriogenesis soma daripada bunga jantan tidak matang empat kultivar *Musa* spp. iaitu. Berangan, Rastali, Mas dan Raja serta mengoptimakan protokol transformasi gen menerusi peledakan mikroprojektil pada bunga jantan tidak matang *Musa* spp. cv. Rastali.

Bunga jantan tidak matang adalah meristem berpotensi tinggi dan bahan yang ideal untuk organogenesis dan embriogenesis soma pada *Musa* spp. Bunga jantan tidak matang bagi empat kultivar *Musa* spp. iaitu. Berangan, Rastali, Mas dan Raja dikultur pada medium MS dimodifikasi yang mengandungi 0, 9, 18 dan 36 µM BAP. Kajian ini menunjukkan peratus induksi meristem tunas adventitius yang tinggi pada medium MS dengan 9 µM BAP bagi kultivar Rastali dan Raja manakala 18 µM BAP bagi kultivar Berangan dan Mas dalam tempoh 4 hingga 8 minggu selepas inisiasi kultur. Regenerasi



pucuk daripada tunas adventitius yang diperoleh daripada bunga jantan tidak matang *Musa* spp. *cv.* Berangan, Rastali, Mas dan Raja dikaji pada medium MS mengandungi BAP berkepekatan 4.5, 9, 18 dan 36 µM selepas empat minggu dikultur dengan pensubkulturan pada setiap minggu. Melalui pemerhatian didapati bahawa 4.5 µM BAP menghasilkan bilangan pucuk tertinggi manakala 36 µM BAP menghasilkan panjang pucuk tertinggi pada semua kultivar yang dikaji selapas empat subkultur. Didapati bahawa subkultur mempengaruhi bilangan min pucuk yang terhasil dengan min bilangan pucuk mencapai 40.33, 5.66, 23.66 dan 19.00 bagi kultivar Rastali, Raja, Berangan dan Mas pada subkultur ke-3 (dari empat subkultur). Min ketinggian pucuk turut meningkat pada setiap kitaran subkultur bagi setiap kultivar.

Gabungan air kelapa (pensterilan turas dan diautoklaf) dengan kepekatan BAP yang telah dikenalpasti paling sesuai untuk induksi tunas adventitius bagi setiap kultivar dari kajian sebelumnya juga telah dikaji. Bilangan min tunas adventitius tertinggi selepas tiga subkultur (17.30) diperoleh pada medium yang mengandungi 50 mlL⁻¹ air kelapa (pensterilan turas) dengan kombinasi BAP pada 9 μ M bagi cv. Rastali dan Raja manakala 18 μ M bagi cv. Berangan dan Mas. Pucuk yang diperoleh daripada bunga jantan tidak matang cv. Berangan, Rastali, Mas dan Raja diakarkan pada medium MS separa kepekatan yang mengandungi 1.0 μ M IBA dan plantlet yang terhasil diaklimatisasi di dalam kebuk tumbesaran.

Kajian histologi dan mikroskopi pengimbas elektron (SEM) telah dijalankan ke atas bunga jantan *Musa* spp. cv. Rastali, yang dikulturkan di dalam rawatan 9 µM BAP,

vii

pada peringkat permulaan kultur dan pada subkultur pertama, kedua dan keempat. Perubahan yang berturutan jelas diperoleh bermula daripada struktur globul (benjolan) sehingga pembentukan tunas adventitius dan akhirnya pengeluaran tunas berganda.

Embriogenesis soma bagi *Musa* spp. cv. Raja telah diperoleh dengan menggunakan bunga jantan tidak matang. Peratus tertinggi pembentukan kalus embriogenik (41.97%) diperoleh pada 13.5 μ M 2,4-D untuk semua posisi kluster bunga yang diuji. Kajian ini menunjukkan sel ampaian embriogenik mempunyai potensi yang tinggi terhadap pembentukan embrio soma. Peratus percambahan embrio soma tertinggi (62.36%) diperoleh pada 0.17 μ M BAP selepas dua minggu dikultur.

Kajian transformasi menunjukkan jarak sasaran 9 cm dan tekanan helium 1350 dan 1550 psi adalah kombinasi yang baik untuk transformasi bunga jantan tidak matang cv. Rastali melalui teknik peledakan mikroprojektil di mana 58.26% dan 57.63% kultur yang bagi setiap kombinasi menunjukkan ekspresi gen GFP yang tinggi.

Keseluruhannya kajian ini menunjukkan bunga jantan tidak matang *Musa* spp. cv. Berangan, Rastali, Mas and Raja sebagai bahan yang paling sesuai untuk regenerasi *in vitro* melalui organogenesis dan embriogenesis soma dan juga untuk transformasi gen menerusi kaedah peledakan mikroprojektil.



ACKNOWLEDGEMENTS

All praises and thanks to almighty Allah, the Most Gracious and Merciful; upon His permission I could complete this thesis.

I would like to express my sincere appreciation to my supervisor and chairman of my supervisory committee, Associate Professor Dr. Maheran Abdul Aziz of the Department of Agriculture Technology, Faculty of Agriculture, Universiti Putra Malaysia- a patient and understanding lady with all intellectual attitudes and kind support beyond her duties at any time even in her tight schedule. Her invaluable guidance, kind advice and help during the course of this research encouraged me to constantly pursue and successfully complete this thesis.

I would also like to express my deep gratitude to my committee member, Mr. Azmi Abdul Rashid, M.Phil., Department of Agriculture Technology, Faculty of Agriculture for his constructive suggestions and his endless enthusiasm to guide and answer my academic questions in a very friendly manner. His invaluable co-operation in completing this thesis is never waved. My gratitude to Associate Professor Datin. Siti Nor Akmar Abdullah, Department of Agriculture Technology, Faculty of Agriculture for her guidance and constructive criticism during the course of this project.

I would also like to express my thanks to my dearest father- Hossein Kashefi, my mother- Akhtar Rokhi, my beloved and patient spouse- Hassan Nazari Moghadam, my little cooperative daughter- Nikta Nazari Moghadam, my brother Amir Kanan Kashefi,



my sister Kiana Kashefi, and my very kind aunt and my dearest grand parents in the hometown- Afsar Rokhi, Akram Shemirani and Ali Rokhi for their encouragement, warm cooperation, moral inspiration during the period of my study.

Sincere appreciation is extended to Associate Professor Dr. Abd Ghani Yunus for the facilities provided on histological study and his guidance, kind assistance as well as Mr. Daud Mustam for his cooperative help in Histology Laboratory, Department of Crop Science, Faculty of Agriculture.

I am also indebted to Associate Professor Dr. Mihdzar Bin Abdul Kadir, the former Head of Department of Agriculture Technology, Faculty of Agriculture for the facilities provided on acclimatization study and help in completion of this thesis.

Finally, special thanks to my lab mates Aini Mohd. Zainol Azlin (my best friend), Beverlien Christine Daiman, Norwaty Bita and Dalila Bahrun for their help and kind cooperation.

Х

I certify that an Examination Committee met on 20th May 2008 to conduct the final examination of Keynoosh Kashefi on her Master of Agriculture Science thesis entitled "Organogenesis, Somatic Embryogenesis and Regeneration of Immature Male Flowers of Banana Cultivars" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Act 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Uma Rani Sinniah, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairperson)

Midhzar Abdul Kadir, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Nor' Aini Mohd. Fadzillah, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Abdul Rahman Milan, PhD

Deputy Director Horticulture Research Centre MARDI (External Examiner)

HASANAH MOHD. GHAZALI, PhD

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Agricultural Science. The members of the Supervisory Committee were as follows;

Maheran Abdul Aziz, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Azmi Abdul Rashid, M.Phil.

Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

Siti Nor Akmar Abdullah, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

AINI IDERIS, PhD

Professor And Dean School of Graduate Studies Universiti Putra Malaysia

Date: 10 July 2008



DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously and is not concurrently submitted for any other degree at UPM or at any other institution.

KEYNOOSH KASHEFI

Date:



TABLE OF CONTENTS

DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xii
LIST OF TABLES	xviii
LIST OF PLATES	xxi
LIST OF FIGURES	XXV
LIST OF ABBREVIATIONS/NOTATIONS	xxvi

CHAPTER

INTRODUCTION		1	
	1.1	Background	1
	1.2	Problem statement	5
	1.3	Objectives of the Study	5
	1.4	Significance of the Study	6

2 LITERATURE REVIEW

2.1	In Vitr	o System	7
2.2	Banana Tissue Culture		8
	2.2.1	Growth Regulators Requirement in Tissue Culture of Banana	8
	2.2.2	Explant Type and Preparation	13
	2.2.3	Somaclonal Variation	14
2.3	Organogenesis		15
	2.3.1	In vitro Shoot Induction	15
	2.3.2	In vitro Root Induction	18
2.4	4 Acclimatization		19
2.5	Somat	ic Embryogenesis	20
	2.5.1	Embryogenic Callus Induction and Cell Suspension	21
		Maturation and Plant Recovery	26
2.6		mical And Micromorphological Study	27
2.7	Plant (Gene Transformation	29

3 MATERIALS AND METHODS

3.1 Plant Regeneration Through Organogenesis From Immature Male 36

xiv

	Flowers Of Banana (Musa Spp.) Cv. Berangan, Rastali, Mas And Raja		
	3.1.1	Induction of Adventitious Bud from Immature Male Flowers of <i>Musa</i> spp. cvs. Berangan, Rastali, Mas and Raja	41
	3.1.2	Shoot Regeneration from the Adventitious Buds of Musa spp.	42
	3.1.3	cvs. Berangan, Rastali, Mas and Raja Effect of Different Coconut Water Preparations in Combination with the Best BAP Treatment on Adventitious Bud Induction from Immature Male Flowers of <i>Musa</i> spp. cvs. Rastali, Raja, Berangan and Mas	43
	3.1.4 3.1.5	In Vitro Root Induction Acclimatization of Plantlets Regenerated from Immature Male Flowers	43 44
3.2		mical And Micromorphological Investigation On Stages Of titious Bud Formation From Male Flowers Of <i>Musa</i> Spp. Cv.	45
	3.2.1	Light Microscopy Scanning Electron Microscopy (SEM)	45 47
3.3	3.3 Plantlet Regeneration Through Somatic Embryogenesis From Immature Male Flowers Of Banana (<i>Musa</i> Spp.) Cv. Raja		48
	3.3.1 3.3.2	Induction of Embryogenic Callus from Immature Male Flowers Establishment of Cell Suspension Culture and Early Somatic Embryo Formation	48 51
	3.3.3	Somatic Embryos Germination and Plantlet Formation	53
3.4		Preliminary Study On Gene Transformation Of Banana (Musa Spp.) Cv. Rastali Via Particle Bombardment	55
	RESU	LTS	61
4.1	Plant 1	Regeneration Through Organogenesis From Immature Male rs Of Banana (<i>Musa</i> Spp.) Cv. Berangan, Rastali, Mas And Raja	61
	4.1.1	Effects of Different Concentrations of BAP on Adventitious Bud Induction from Immature Male Flowers of <i>Musa</i> spp. cv.	61
	4.1.2	Berangan, Rastali, Mas and Raja Regeneration of Shoots from the Adventitious Buds Derived from Immature Male Flowers of Banana (<i>Musa</i> spp.) Cultivars	69
	4.1.3	Berangan, Rastali, Mas and Raja after Four Subcultures Effect of Different Coconut Water Preparation with BAP on Adventitious Bud Induction from Immature Male Flowers of <i>Musa</i> spp. cvs. Rastali, Raja, Berangan and Mas	75

4

Musa spp. cvs. Rastali, Raja, Berangan and Mas4.1.4 Rooting of Shoots Derived from Immature Male Flowers of 78



Musa spp. cvs. Berangan, Rastali, Mas and Raja

- 4.1.5 Acclimatization of Plantlets Regenerated from Immature Male 81 Flowers of cvs. Rastali, Raja, Berangan and Mas
- 4.2 Anatomical And Micromorphological Investigation On Stages Of 87 Adventitious Bud Formation From Male Flowers Of *Musa* Spp. Cv. Rastali
- 4.3 Plantlet Regeneration Through Somatic Embryogenesis From Immature 100 Male Flowers Of *Musa* Spp. Cv. Raja
 - 4.3.1 Embryogenic Callus Induction and Somatic Embryo Formation 100 from Immature Male Flowers of *Musa* spp. cv. Raja
 - 4.3.2 Establishment of Embryogenic Cell Suspension from the 107 Embryogenic Complex
 - 4.3.3 Somatic Embryo Maturation in Cell Suspension 112
 - 4.3.4 Somatic Embryo Germination and Plantlet Formation 114
- 4.4 Preliminary Study On Gene Transformation Of *Musa* Spp. Cv. 119 Rastali Using Particle Gun Bombardment Technique

5 DISCUSSION

122

- 5.1 Plant Regeneration Through Organogenesis From Immature Male 122 Flowers Of Banana (*Musa* Spp.) Cv. Berangan, Rastali, Mas And Raja
 - 5.1.1 *In vitro* Shoot Regeneration 122
 - 5.1.2 *In vitro* Rooting and Plantlet Acclimatization 128
- 5.2 Anatomical And Micromorphological Investigation On Stages Of 130 Adventitious Bud Formation From Male Flowers Of *Musa* Spp. Cv. Rastali
- 5.3 Plantlet Regeneration Through Somatic Embryogenesis From Immature 133 Male Flowers Of *Musa* Spp. Cv. Raja
 - 5.3.1 Embryogenic Callus Induction and Somatic Embryo Formation 133 from Immature Male Flowers of *Musa* spp. cv. Raja
 - 5.3.2 Establishment of Embryogenic Cell Suspension from 135 Embryogenic Complex
 - 5.3.3 Somatic Embryo Maturation and Germination 137
- 5.4 Preliminary Study On Gene Transformation Of *Musa* Spp. Cv. 139 Rastali Using Particle Gun Bombardment Technique

6 GENERAL DISCUSSION AND CONCLUSION 141



150
178
201



LIST OF TABLES

Table		Page
4.1	Adventitious bud induction from immature male flowers of Musa spp.	63
	cv. Rastali in different BAP treatments over subculture cycles.	
4.2	Adventitious bud induction from immature male flowers of Musa spp.	63
	cv. Raja in different BAP treatments over subculture cycles.	
4.3	Adventitious bud induction from immature male flowers of Musa spp.	65
	cv. Berangan in different BAP treatments over subculture cycles.	
4.4	Adventitious bud induction from immature male flowers of Musa spp.	66
	cv. Mas in different BAP treatments over subculture cycles.	
4.5	Adventitious buds induction from immature male flowers of Musa spp.	67
	cv. Rastali, Raja, Berangan and Mas on different BAP levels after eight	
	weeks of culture.	
4.6	Total number of shoot proliferated from adventitious buds of Musa spp.	70
	cvs. Rastali, Raja, Berangan and Mas on MS medium with modified	
	vitamins supplemented with different levels of BAP from four weekly	
	subcultures.	
4.7	Height of shoots attained from adventitious buds of Musa spp. cvs.	71
	Rastali, Raja, Berangan and Mas on MS medium with modified vitamins	
	supplemented with different levels of BAP after 4 weeks of culture.	
4.8	Multiple shoot proliferation from adventitious buds of Musa spp. cvs.	73
	Rastali, Raja, Berangan and Mas on MS medium with modified vitamins	
	supplemented with $4.5\mu M$ BAP over subculture cycles.	



- 4.9 Height of shoots attained from adventitious buds of *Musa* spp. cvs. 73
 Rastali, Raja, Berangan and Mas on MS medium with modified vitamins supplemented with 4.5μM BAP over subculture cycles.
- 4.10 Effects of different coconut water preparations with BAP on mean 76 number of adventitious buds produced per explant in different banana cultivars after three subcultures.
- 4.11 Effects of different coconut water preparations on mean number of 77 adventitious bud produced per explant over subculture cycle for all banana cultivars.
- 4.12 Number of roots produced per explant in banana cultivars over 5 weeks 79 of culture.
- 4.13 Root length (cm) attained on shoots derived from male inflorescences of 79 banana cultivars at week 5 of culture.
- 4.14 The effects of different potting media on survival rate, number of leaves 84 produced and plant height (cm) attained at week 6 of acclimatization in cv. Rastali.
- 4.15 The effects of different potting media on survival rate, number of leaves 84 produced and plant height (cm) attained at week 6 of acclimatization in cv. Raja.
- 4.16 The effects of different potting media on survival rate, number of leaves 85 produced and plant height (cm) attained at week 6 of acclimatization in cv. Berangan.



- 4.17 The effects of different potting media on survival rate, number of leaves 85 produced and plant height (cm) attained at week 6 of acclimatization in cv. Mas.
- 4.18 Effect of different concentrations of 2,4-D on embryogenic callus 103 formation from cv. Raja (observed after 6 months).
- 4.19 Effect of different flower hand positions on embryogenic callus 104 formation from cv. Raja (observed after 6 months).
- 4.20 Interaction between 2,4-D concentrations and flower hand positions on 105 percentage of embryogenic callus formation from cv. Raja (observed after 6 months).
- 4.21 Percentage of somatic embryo germination on Cote *et al.* (1996) 117 medium supplemented with different concentrations of BAP after four weeks of culture.
- 4.22 Interaction between the different helium pressures (psi) and target 120 distances on GFP gene expression in immature male flowers of *Musa* spp. cv. Rastali 48 hours after bombardment.



LIST OF PLATES

Plate		Page	
3.1	Male inflorescence of banana with a fruit bunch.	38	
3.2	Close up view of A) An opened immature male flower bud and B) A	39	
	horizontal section of a banana male flower.		
3.3	Preparation, surface sterilization and culture of floral buds.	40	
3.4	Flower hands excised from a male inflorescence of cv. Raja.	51	
3.5	Sectioned immature male flower buds of Musa spp.cv. Rastali	57	
	compactly placed in the centre of each 9 cm disposable petridish		
	containing MS medium supplemented with 9 µM BAP.		
4.1	Immature male flower cultures of Musa spp. cv. Rastali.	64	
4.2	Response of immature male flowers of banana cvs. Rastali, Berangan,	68	
	Raja and Mas on MS medium with BAP after four subcultures.		
4.3	Shoot regeneration from adventitious buds derived from male	72	
	inflorescences of banana cultivars Rastali, Raja, Berangan and Mas after		
	2 weeks (second subculture) of culture.		
4.4	Shoot regeneration from adventitious bud derived from male	74	
	inflorescences of banana cultivars Rastali, Raja, Berangan and Mas after		
	4 weeks (fourth subculture) of culture.		
4.5	Effects of different coconut water preparations in combination with BAP	77	
	on adventitious bud production in different banana cultivars after three		
	weeks of culture.		

4.6 Rooting of shoots of banana cv. Rastali (with long and thick roots plus 80



root hairs), cv. Raja (with short and weak roots), cv. Berangan (long and thin roots) and cv. Mas (with hairy moderate roots) at week 3 of culture.

- 4.7 Root development on shoots of banana cultivars at week 5 of culture. 81
- 4.8 Growth of plantlets in sand + peat moss in the growth chamber. 83
- 4.9 Performance of plantlets of banana cultivars after six weeks of 86 acclimatization in different potting media.
- 4.10 Stages of adventitious bud induction from immature male flowers of 88*Musa* spp. cv. Rastali on MS medium with 9μM BAP.
- 4.11 Immature male flower bud of *Musa* spp. cv. Rastali at the initial culture 90 stage.
- 4.12 Immature male flower bud of *Musa* spp. cv. Rastali at week 2 of culture 91 (first subculture) on MS medium supplemented with 9 μM BAP.
- 4.13 Close up view of a flower hand of a male flower bud of *Musa* spp. cv. 92 Rastali at second subculture (week 4) on MS medium supplemented with 9 μM BAP.
- 4.14 Histological view of a flower finger of *Musa* spp. cv. Rastali on 93MS medium with 9 μM BAP.
- 4.15 Scanning electron microscopic view of a flower hand of *Musa* spp. cv. 94Rastali on MS medium with 9 μM BAP.
- 4.16 Close up view of a flower hand of a male flower bud of cv. Rastali at 95 fourth subculture (week 8) on MS medium with 9 μM BAP.
- 4.17 Histological section showing an undulating zone with furrows 96 separating the tiny meristems at the base of a flower finger (fg) in a



flower hand at an early stage.

- 4.18 Scanning electron microscopic view of the adventitious bud clusters 97 induced at the base of a fully-enlarged flower finger (green arrow) in a flower hand of an immature male flower bud of *Musa* spp. cv. Rastali at fourth subculture on MS medium with 9 μM BAP.
- 4.19 Histological view of stages of adventitious shoot induction from 98 immature male flowers of *Musa* spp.cv. Rastali.
- 4.20 Scanning Eelctron Microscopic view of adventitious shoot induction 99 stages from immature male flowers of *Musa* spp. cv. Rastali.
- 4.21 Callus induction from immature male flowers of *Musa* spp. cv. Raja on 101 MS medium supplemented with 1 mgL⁻¹ biotin, 1 mgL⁻¹ IAA, 1 mgL⁻¹ NAA and different concentrations of 2,4-D.
- 4.22 Abnormal callus and somatic embryos formation from immature male 106 flowers of *Musa* spp. cv. Raja on MS medium supplemented with 1 mgL⁻¹ biotin, 1 mgL⁻¹ IAA, 1 mgL⁻¹ NAA and different concentrations of 2,4-D.
- 4.23 Embryogenic cell suspension in Becker *et al.* (2001) cell suspension 109 medium.
- 4.24 Microscopic (Inverted microscope: ZEIZZ/ AXiovert 135) view of 110
 Musa spp. cv. Raja embryogenic cells at different culture stages in a medium based on Becker et al. (2001).
- 4.25 Embryogenic cell suspension of immature male flowers of *Musa* spp. cv 112Rastali in Georget *et al.* (2000) cell suspension medium.



4.26	Stages of somatic embr	yo maturation in Musa spp	. cv. Raja.	114

- 4.27 Somatic embryo germination of *Musa* spp. cv. Raja on Cote *et al.* (1996) 116 germination medium.
- 4.28 Somatic embryo germination and plantlet development in *Musa* spp. cv. 118 Raja.
- 4.29 Confirmation of green-fluorescent protein (GFP) gene expression in 121 immature male flowers under the fluorescent microscope with UV light excitation.

