

UNIVERSITI PUTRA MALAYSIA

SYNTHESIS AND CHARACTERIZATION OF SEMISYNTHETIC METALLOTHERMOLYSIN

SYARAJATUL ERMA KHALID

FS 2008 17



SYNTHESIS AND CHARACTERIZATION OF SEMISYNTHETIC METALLOTHERMOLYSIN

By

SYARAJATUL ERMA KHALID

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science.

April 2008



SYNTHESIS AND CHARACTERIZATION OF SEMISYNTHETIC METALLOTHERMOLYSIN.

SYARAJATUL ERMA KHALID

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA

2008



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

SYNTHESIS AND CHARACTERIZATION OF SEMISYNTHETIC METALLOTHERMOLYSIN

By

SYARAJATUL ERMA KHALID

April 2008

Chairman : Associate Professor Mohd. Basyaruddin Abdul Rahman,

PhD

Faculty : Science

Modification of native enzyme has gone through various strategies and evolution of the process itself. Thermolysin (TLN) from Bacillus thermoproteolyticus rokko was modified semisynthetic as а metallothermolysin which comprises enzyme, ligand and metal ion. Ligands used were benzamidine (BEN), 1,10-phenanthroline (PHN), paminobenzamidine (PBZ) and ethanolamine (ETA). The metal ions chosen were magnesium (Mg²⁺), zinc (Zn²⁺), calcium (Ca²⁺) and nickel (Ni²⁺). The semisynthetic metallothermolysin activities were evaluated on hydrolysis reaction of azocasein. Among the four ligands, complex of TLN-PBZ showed the highest specific activity (2219.5 Unit per mg (U/mg)) at optimum PBZ concentration of 0.6 mM. The study followed by the attachment of Mg²⁺ to TLN-PBZ complex which gave the best specific activity compared to other metal ions (39406.4 U/mg). The optimum concentration of Mg²⁺ was found



best at 0.08 mM. Several parameters were also investigated such as studies on effect of pH, temperature, time course and thermostability. As a result, the semisynthetic metallothermolysin maintained at pH surrounding of 7.0 in tris-HCI buffer and found optimum at 80°C for reaction up to 3 hours (96.7% of relative activity). For thermostability test, the semisynthetic metallothermolysin can retain its activity up to 90% at pre-heated temperature of 80°C.

Electronic absorption like the UV/Visible (UV/Vis) and UV/Fluorescence spectrophotometer and Circular Dichroism (CD) spectropolarimetry method were used to characterize the optical properties of metallothermolysin. In UV/Vis spectrophotometer, the binding of PBZ to TLN curve caused a bathocromic shift (λ_{max} from 279 nm to 274 nm) and became hypsochromicsm (λ_{max} from 274 nm to 272 nm) with the additional of Mg²⁺. Changes in UV/Vis were also supported by UV/Fluorescence, when changes happened to the emission characteristic of TLN-PBZ spectrum (373.2 nm) and the spectrum continues to shift (374.0 nm) for TLN-PBZ-Mg. The CD spectropolarimetry suggested some changes of α helix and β sheet at far UV molar ellipticity readings with decreased of α helix from 37% (TLN) to 20.6% (TLN-PBZ) and then to 19.8% (TLN-PBZ-Mg). Meanwhile, a further decrease of β sheet from 32.6% (TLN) to 18.7% (TLN-PBZ) and then to 11.0% (TLN-PBZ-Mg) was also observed.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Master Sains

SINTESIS DAN PENCIRIAN METALLOTHERMOLYSIN SEMISINTETIK

Oleh

SYARAJATUL ERMA KHALID

April 2008

Pengerusi : Profesor Madya Mohd. Basyaruddin Abdul Rahman, PhD

Fakulti : Sains

Modifikasi enzim telah melalui pelbagai strategi dan mengalami evolusi tersendiri. Thermolysin (TLN) merupakan sejenis enzim yang telah diekstrak dari *Bacillus thermoproteolyticus rokko*. Ia telah diubahsuai menjadi metalloenzim semisintetik yang terdiri daripada gabungan enzim, ligan dan ion logam. Beberapa ligan yang digunakan adalah benzamidina (BEN), 1,10-phenantrolina (PHN), p-aminobenzamidina (PBZ) dan etanolamina (ETA). Ion-ion logam yang terlibat pula adalah magnesium (Mg²⁺), zink (Zn²⁺), kalsium (Ca²⁺) dan nikel (Ni²⁺). Aktiviti metallothermolysin semisintetik diperolehi dari tindak balas hidrolisis azocasein. Antara empat jenis ligan tersebut, kompleks TLN-PBZ menunjukkan aktiviti spesifik yang tertinggi (2219.5 U/mg) pada kepekatan optimum PBZ, 0.6 mM. Ujikaji diteruskan dengan penambahan Mg²⁺ pada kompleks TLN-PBZ yang menghasilkan aktiviti spesifik tertinggi berbanding ion logam lain (39406.4 U/mg). Kepekatan optimum Mg²⁺ adalah 0.08 mM. Beberapa parameter telah diuji



kesan pH, suhu, masa tindak balas dan kestabilan suhunya. Sebagai keputusannya, metallothermolysin semisintetik ini dapat mengekalkan persekitaran dalam larutan penimbal tris-HCl pada pH 7.0 manakala tindak balas optimumnya adalah pada 80°C selama 3 jam (peratusan aktiviti relatif sebanyak 96.7 %). Metallothermolysin semisintetik ini stabil suhu walaupun melalui pra pemanasan pada suhu 80°C di mana ia mampu bertindak balas dengan kadar 90 %.

Analisis spektroskopi seperti Ultra lembayung boleh nampak (UV/Vis), Ultra lembayung fluoresen (UV/*Fluorescence*) spektrofotometer dan *Circular Dichroism* (CD) spektropolarimeter digunakan bagi tujuan pencirian aset optikal metalloenzim semisintetik. Keputusan UV/Vis menunjukkan berlaku anjakan pada bacaan panjang gelombang apabila perlekatan PBZ ke TLN berlaku (λ_{max} dari 279 ke 274 nm) dan bersifat hipokromik (λ_{max} dari 274 nm ke 272 nm) apabila Mg²⁺ ditambah. Perubahan bacaan UV/Vis disokong keputusan bacaan UV/*Fluorescence* yang menunjukkan perubahan bacaan panjang gelombang bagi spektrum TLN-PBZ (373.2 nm) dan anjakan spektrum terus berlaku (374.0 nm) bagi TLN-PBZ-Mg. Keputusan CD spektropolarimetri mengusulkan berlakunya perubahan struktur protein α heliks dan β *sheet* di mana bacaan unit molar eliptisiti α heliks menurun dari 37.0 % (TLN), ke 20.6 % (TLN-PBZ) dan seterusnya 19.8 % (TLN-PBZ-Mg). Bagi struktur β *sheet* penurunan dari 32.6 % (TLN) kepada 18.7 % (TLN-PBZ) sehingga ke 11.0 % (TLN-PBZ-Mg).

ACKNOWLEDGEMENTS

Alhamdulillah, praises to Allah s.w.t. for giving me the strength to endure all problems and complete this study.

I wish to express my sincere appreciation and gratitude to my supervisor, Associate Professor Dr. Mohd Basyaruddin Abdul Rahman for his patience and persistent encouragements. Thank you to my supervisory committee, Professor Dr. Mahiran Basri and Professor Dr. Abu Bakar Salleh for their great concern, advices and invaluable assistance from the beginning till the end of this study.

Thank you also to the staff members of the Department of Chemistry who were so helpful and cooperative in many ways during the course of the study.

I wish to thank all my friends and members of Lab 401, Kak Salina, Kak Yati, Ita, Redzuan, Pei Sin, Lam, Us, Mona, Azizah, Nora, Shie Ling, Hasmah and Casey for the friendship and for making my stay in UPM a memorable one with many sweet memories and experiences. Thank you for being friends in need.

My deep expression was also extended to Biochemistry lab mates especially, Kak Ina, Kak Ain, Aiman, Ghani, Leow, Kok Whye, Ropandi and the others for their valuable help.



Not forgotten my colleague in Mardi, Ezy, Dayana, Nisa, Sabeetha and Kak Ju, thank you for your help and support.

Finally, my deepest appreciation goes to my parents, papa and mama, for their never-ending moral and constant support during my studies. Not forgetting, my special thanks to my husband, Wan Ahmad Marzuki B. Wan Ahmad for his patience and to my son, Wan Muhammad Syamim, you are my inspiration.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd. BasyarudDin Abd. Rahman, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Chairman)

Mahiran Basri, PhD

Professor Faculty of Science Universiti Putra Malaysia (Member)

Abu Bakar Salleh, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

AINI IDERIS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 10 July 2007



DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that is has not been previously, and is not concurrently submitted for any other degree at UPM or at any other institution.

SYARAJATUL ERMA BINTI KHALID

Date: 24 June 2008



viii

TABLE OF CONTENTS

ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENT	V
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xiii
LIST OF FIGURES	xiiv
LIST OF EQUATIONS	xvi
LIST OF ABBREVIATIONS	xvii

CHAPTER

1	INT	RODU	ICTION	1
2	LIT	ERATL	JRE REVIEW	5
	2.1	Prote	ases	5
	2.2	Metal	lo Protease (EC number 3.4.24)	7
		2.2.1	Thermolysin	12
	2.3	Semi	synthetic metalloenzyme	15
	2.4	Ligan	ds	17
		2.4.1	Types of ligands	19
	2.5	Metal	lions	21
		2.5.1	Type of metal ions	22
	2.6	Struc	tural Spectroscopic Studies	23
		2.6.1	UV/Visible	24
		2.6.2	UV/Fluorescence	25
		2.6.3	Circular Dichroism (CD) Spectropolarimetry	28
3	MA	TERIA	LS AND METHODS	31
	3.1	Cherr	nicals	31
	3.2	Metho	ods	33
		3.2.1	Purification of Protease	33
		3.2.2	Protease Assay	34
		3.2.3	Protein Determination	35
			Characterization and optimization of the enzyme	35
		3.2.5	Synthesis of the Semisynthetic	
			Metallothermolysin	37
		3.2.6	Electronic Absorption Measurement	39
4	RE	SULTS	S AND DISCUSSIONS	
	4.1	Dete	rmination of the purity of Thermolysin	42
	4.2	Syntl	hesis of Thermolysin-ligand	44

Page



	4.2.1	Optimum Activity Based on	
		p-Aminobenzamidine (PBZ) Concentration	51
4.3	Synth	esis of Thermolysin-Ligand- Metal	56
	4.3.1	Optimum Activity based on Magnesium (Mg)	
		Concentration	62
4.4	Chara	acterization of Metallothermolysin	66
	4.4.1	Optimal pH	66
	4.4.2	Optimal Temperature and Time Course Study of	
		the Selected Temperature	69
	4.4.3	Time Course Study	72
		Thermostability	74
4.5	Struct	tural Studies	76
	4.5.1	Measurements of UV/Vis	77
	4.5.2	UV/Fluorescence	83
	4.5.3	Circular Dichroism (CD) spectra analysis	88
5 CO	NCLU	SION AND RECOMMENDATION	92
5.1	Concl	lusions	92
5.2	Reco	mmendations	94
REFER		S	96
APPEN	DICES	8	106
BIODA	TA OF	THE STUDENT	111



LIST OF TABLES

Table		Page
1	Hard/soft classifications for ligand and cation	18
2	Structure of chemical ligands involved in synthes of semisynthetic metallothermolysin.	is 20
3	Wavelength ranges	29
4	Purification table of TLN	42
5	Purification table of crude (C) and pooled samples (S) of TLN-PBZ, TLN-BEN, TLN-ETA and TLN-PHN using the Sephadex G-50 gel filtration chromatography	48
6	Final E _{docked} (∆G) for different ligands at four largest pockets in thermolysin	54
7	Purification table of crude (C) and pooled samples (S) of TLN-PBZ-Mg, TLN-PBZ-Ca, TLN- PBZ-Ni and TLN-PBZ-Zn using the Sephadex G-50 gel filtration chromatography	59
8	Ratio values (%) calculated from the CD spectropolarimeter	91



LIST OF FIGURES

Figure		Page
1	Hydrolysis reaction of protein catalysed by protease	8
2	Zinc Protease catalytic logic (a) and (b)	9
3	Catalytic mechanisms of metalloenzyme where the active site Zn ²⁺ to coordinate and activate attacking water molecule	11
4	Ribbons representation of thermolysin (1KEI.pdb)	13
5	The active site of thermolysin enzyme	14
6	(a) A π - π * transition of a C–C double bond, with the lower energy π state. (b) A π – σ * transition of a C–C double bond with the same lower energy electron configuration	27
7	SDS-PAGE test. The commercial thermolysin and the purified thermolysin showed single band	43
8	Elution graph of thermolysin enzyme using Sephadex G-50 as matrix in gel filtration chromatography column	44
9	Total unit activity (U) versus time (hours) of bonding between TLN with four selected ligands 46	
10	Total activities and total proteins of newly modified TLN-ligand complexes. (Relative protein % are shown) 50
11	Concentrations (mM) versus enzyme activity (U/ml) graph showed the optimum concentration for PBZ was 0.6 mM in 20 mM tris-HCI buffer pH 7.0. The reaction was at 37 °C	52
12	Visualization of thermolysin structure with four main pockets determined from CASTp: Binding site- pocket 48 (Cyan), Pocket 47 (Green), pocket 46 (Blue) and pocket 45 (Red)	55
13	Visualization on PBZ docked to pocket 45 in thermolysin structure. PBZ docked onto pocket 45 (left) and inset picture of PBZ docked onto pocket 45 (right)	56



14	Total activities and proteins of TLN-PBZ-metal. (Relative protein % shown)	60
15	Concentrations (μM) versus enzyme activity (U/mI) grap showed the optimum concentration for Mg was at 0.08 mM in 20 mM tris-HCI buffer pH 7.0. The reaction was at 37 °C	h 64
16	Visualization on Mg ²⁺ and PBZ docked to pocket number 45 in thermolysin structure	65
17	Effect of pH towards relative activity of TLN, TN-PBZ and TLN-BZ-Mg in pH 4.0, 5.0, 6.0, 7.0, 8.0, 9.0 and 10.0	d 67
18	Effect of temperature towards relative activity of TLN, TLN-PBZ and TLN-PBZ-Mg. Reactions were performed in tris-HCI buffer, pH 7.0 for 30 minutes	70
19	Effect of temperature towards relative activity of TLN, TLN-PBZ and TLN-PBZ-Mg. Reactions were performed in tris-HCI buffer, pH 7.0 for 1 hour	71
20	Time course towards relative activity of TLN, TLN-PBZ and TLN-PBZ-Mg. Reaction were performed triplicate in tris-HCI buffer, pH 7.0 at optimum temperatu of each samples (80°C: TLN-PBZ-Mg; 60°C: TLN-PBZ; 70°C: TLN)	ire 73
21	Effect of pre-incubation temperature towards relative activity of TLN,TLN-PBZ and TLN-PBZ-Mg	75
22	UV/Vis absorption spectrum of pH 7.0 phosphate buffer without PBZ (dotted line) and 0.2 mM PBZ(solid line) between 200 and 500 nm at room temperature. Molar absorption coefficients, ε value (M ⁻¹ cm ⁻¹) (italicized)	78
23	UV/Vis spectra of TLN and TLN-PBZ in 20mM phosphate buffer pH 7.0 at room temperature: (i) spectrum of TLN (0.2mM); (ii) spectrum of TLN-PBZ (0.2mM)	9 80
24	PBZ spectra quenching by Mg in 20mM phosphate buffe pH 7.0 at room temperature. UV-vis spectra of PBZ (i) and PBZ-Mg (ii)	er 82



25	PBZ quenching by Mg in TLN-PBZ (in 20mM phosphate buffer pH 7.0 at room temperature). UV/Vis spectra of (i) TLN-PBZ and (ii) TLN-PBZ in presence of 0.08 mM Mg	83
26	Fluorescence spectra of TLN , TLN-PBZ and TLN-PBZ-Mg in 20 mM phosphate buffer pH 7.0 at 20°C	85
27	Far UV CD spectra of TLN (red), TLN-PBZ (green) and TLN-PBZ-Mg (blue)	89

LIST OF EQUATIONS

EquationPage1Molar absorptivity (M⁻¹ cm⁻¹)262Left and right-circularly polarized light283Molar ellipticity294Determination of Protease Activity35



LIST OF ABBREVIATIONS

Enzyme

Protease X from Bacillus Thermoproteolyticus rokko	TLN
(Thermolysin enzyme)	

Ligands

p-Aminobenzamidine	PBZ
Benzamidine	BEN
Ethanolamine	ETA
1, 10- phenanthrolin e	PHN

Metals/ Metal ions

Cd
Ca ²⁺
Cr
Cu
Fe
K
Li
Mg ²⁺
Mn
Na
Ni ²⁺
Zn ²⁺



Amino Acids

Alanine	Ala
Asparagine	Asn
Aspartate	Asp
Glutamine	Glu
Glysine	Gly
Histidine	His
Isoluecine	lle
Leucine	Leu
Lysine	Lys
Methionine	Met
Serine	Ser
Tyrosine	Tyr
Valine	Val
Adenosine triphosphate	ATP

Spectrocopy Instruments

Circular Dichroism	CD
Ultra-violet/ Fluoresence	UV/Fluo
Ultra-violet/Visible	UV/Vis



Units

Absorbance	[A]
Centimeter	cm
Dalton	Da
Gram	g
Kilo-dalton	kDa
Liter	L
Microgram	μg
Microliter	μl
Mililiter	ml
Miligram	mg
Molar	Μ
Molar absorptivity	3
Molar ellipticity	θ
Optical Density	OD
Temperature	°C
Unit	U
Wavelength	nm



I certify that an Examination Committee has met on 11 April 2008 to conduct the final examination of name of Syarajatul Erma Binti Khalid on her Master of Science thesis entitled "Synthesis and Characterization of Semisynthetic Metallothermolysin" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the Master of science.

Members of the Examination Committee were as follows:

Kamaliah Sirat, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Chairman)

Abdul Halim Abdullah, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Johari Ramli, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Wan Azlina Ahmad, PhD

Professor Faculty of Science Universiti Teknologi Malaysia (External Examiner)

HASANAH MOHD. GHAZALI, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia.

Date: 26 June 2008



CHAPTER 1

INTRODUCTION

Enzymes as biocatalyst are in high demand by the industrial field for their high rates and high reaction specificity and stereoselectivity. Enzymes can accomplish the reactions at mild pH, temperatures and pressures, thereby it consumes less energy. As enzymes are non-toxic, it minimizes problem of downstream waste and by-product disposal in ecologically acceptable processes.

Native enzyme nowadays has gone through evolution where many modifications have been done to fulfill these demands (Abdul Rahman, 1993). One of the modification types is by synthesizing a semisynthetic enzyme. Semisynthetic enzyme refers to an artificial enzyme that was developed at a define site with cofactor or new functional group for its novel properties and unique features. It consists of enzyme, ligand and metal (Rawling and Barrett, 1995).

In this regards, isolated thermolysin from *Bacillus thermoproteolyticus rokko* was selected to be modified to produce a new enzyme known as the semisynthetic metalloenzyme. Thermolysin is known for its high thermostability and many studies have been done by researchers from all over the world (Boonyaras *et al.*, 2000).



According to Ory *et al.*, (1998), an enzyme has regions on its surface where small molecule or ion can bind. Some binding sites on the surface of enzymes may allow binding but only to a limited range of the chemical compounds. The design for development of semisynthetic enzyme was based on the use of protein pockets that can accommodate ligand as an intermediate between the pockets at the surface of the enzyme and metal. According to Conn *et al.*, (1987), approximately one-third of known enzymes has metals as part of their structure, which requires metals to be added for activity or is further activated by metals. Several additions of ligand and metal were highlighted in this study to observe the best complex synthesized that was capable in enhancing or inhibiting the enzyme reaction.

Recently, protein engineering and chemical modification has become a successful valuable tool for creating or improving protein function for practical uses. Therefore introducing cofactors or other reactive moieties into proteins provides enormous flexibility for the design of semisynthetic catalysts that could be employed for a variety of purposes especially to enhance its reaction activity (Distefano and Davies, 1997).

Understanding the structural and functional significance of these ligand and metal effects requires a specialized array of sophisticated instrumentation and techniques as well as the expertise to use them. It is only through a detailed understanding of structure and function that enzymes can be selected or redesigned to perform industrially relevant catalysis (Kazlauskas, 2000). Owing to its inherent sensitivity, simplicity and to some extent



selectivity, UV/Visible (UV/Vis) and UV/Fluoresence spectroscopy were among the selected spectroscopy techniques for more valuable structural proposal (Donald *et al.*, 2001). Circular Dichroism (CD) spectropolarimetry was also used for prediction of the secondary structure of a protein that was modified.

This research focused on developing positive biocatalysts for a variety of purposes especially in pharmaceutical and chemical industries. Therefore the objectives of this study are:

- To design and synthesize metalloenzyme as biocatalyst in bio-based industries.
- To study the characterization and optimization of the modified enzyme.
- To evaluate the activity of the novel semisynthetic metallothermolysin through hydrolysis of azocasein.
- To analyze the semisynthetic metalloenzyme structure using modern spectroscopy.

In order to fulfill the above objectives, the native thermolysin had gone through screening before modification. The proteolytic activity was determined by using azocasein as the substrates. Identified as hydrolases



enzyme which cleave peptide bond, thermolysin catalyzed amide (peptide) bond hydrolysis in protein or peptide substrates.

Parameters involved were the optimum pH, temperature, reaction time and thermostability. These screening results helped us to determine the suitable environment of the native enzyme. Modification steps of the native enzyme were held by mixing the enzyme with ligands and metal ions (one at a time) to form a couple of protein complexes. Then these protein complexes were purified and assayed. The total activity of each complex had been compared among the protein complexes and native enzyme activity. The protein complex that yields highest total activity was chosen for further investigation. The chosen complex was characterized and optimized for its pH surrounding, temperature and thermostability.

This was followed by confirming its structure with an electronic probe such as UV/Vis, UV/Fluorescence spectroscopy and CD spectropolarimeter. The use of UV/Vis proved to us on how different molecules absorbed spectrum showed a number of absorption bands corresponding to structural groups within the molecule at different wavelengths. The same absorbance understanding goes to UV/Fluorescence spectroscopy that had been used and applied to study the fundamental physical processes of molecules and one of them was in structure–function relationships and interactions of biomolecules such as proteins and nucleic acids. CD spectropolarimeter on the other hand will predict protein secondary structure by obtaining information from the UV region of the spectrum.

