



**UNIVERSITI PUTRA MALAYSIA**

**ANALYSIS OF TRANSCRIPTOMIC CHANGES IN OIL PALM (*Elaeis guineensis* Jacq.) ROOT UPON INOCULATION WITH *Bacillus sphaericus* UPMB10 USING cDNA MICROARRAY**

**LIM KOK ANG**

**FBSB 2008 4**



**ANALYSIS OF TRANSCRIPTOMIC CHANGES IN OIL PALM (*Elaeis guineensis*  
Jacq.) ROOT UPON INOCULATION WITH *Bacillus sphaericus* UPMB10 USING  
cDNA MICROARRAY**

**By**

**LIM KOK ANG**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**June 2008**



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**ANALYSIS OF TRANSCRIPTOMIC CHANGES IN OIL PALM (*Elaeis guineensis* Jacq.) ROOT UPON INOCULATION WITH *Bacillus sphaericus* UPMB10 USING cDNA MICROARRAY**

By

**LIM KOK ANG**

**June 2008**

**Chairman: Ho Chai Ling, PhD**

**Faculty: Faculty of Biotechnology and Biomolecular Sciences**

*Bacillus sphaericus* UPMB10 is a plant growth promoting bacteria (PGPB) which enhances plant growth by acting as bioenhancer and biofertilizer. An oil palm cDNA microarray containing 2224 cDNA probes from root (partially derived from cDNA clones generated in this study), 828 from vegetative meristem and 696 from zygotic embryo was generated to investigate the transcriptomic changes in two-month-old *in vitro* oil palm (*Elaeis guineensis* Jacq.) (Deli x Yangambi) roots after inoculation with *B. sphaericus* UPMB10 for 120 h. This study was initiated with the generation of 1824 expressed sequenced tags (ESTs) from the roots of oil palm. A total of 1173 tentative unique genes (TUGs) were assembled from 1566 ESTs with readable sequences. However, only 984 TUGs showed significant matches (*E*-value less than  $10^{-5}$ ) to the non-redundant protein database in the GenBank and they were further divided into 13 groups based on their putative functions. Subsequent microarray result showed that 151 and 125 transcripts were significantly up- and down regulated, respectively in the



roots of oil palm inoculated with *B. sphaericus* UPMB10. Although transcripts involved in protein synthesis were increased and the expression level of auxin responsive genes were altered in *B. sphaericus* UPMB10-inoculated oil palms, there was no conclusive result to support the presence of auxin secreted by *B. sphaericus* UPMB10 in the medium. Despite the capability of *B. sphaericus* UPMB10 to fix atmospheric nitrogen, 120 h might not be sufficient for it to establish efficient nitrogen fixation to relieve possible N deficiency in oil palms. The plant-microbe interaction might also have alerted the defense system that led to the up-regulation of transcripts related to synthesis of hydrogen peroxide in oil palm. Verification of microarray result by real time RT-PCR showed that nine out of 11 candidate genes (81.8%) were consistent in their expression patterns. In conclusion, this study has provided a brief understanding of the transcriptomic changes in the oil palm roots inoculated with *B. sphaericus* UPMB10.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**ANALISASI PERUBAHAN TRANSKRIPTOMIK DI AKAR KELAPA SAWIT  
(*Elaeis guineensis* Jacq.) SELEPAS DIINOKULASI DENGAN *Bacillus  
sphaericus* UPMB10 DENGAN MENGGUNAKAN MIKROATUR cDNA**

Oleh

**LIM KOK ANG**

**Jun 2008**

**Pengerusi: Ho Chai Ling, PhD**

**Fakulti: Fakulti Bioteknologi dan Sains Biomolekul**

*Bacillus sphaericus* UPMB10 merupakan bakteria penggalak pertumbuhan pokok (PGPB) yang boleh mempercepatkan pertumbuhan tumbuhan dengan berfungsi sebagai penggalak biologi dan baja biologi. Satu mikroatur cDNA kelapa sawit yang mengandungi 2224 prob cDNA daripada akar (sebahagiannya berasal daripada klon cDNA yang dijanakan dalam kajian ini), 828 daripada meristem vegetatif, dan 696 daripada embrio zigot telah dijanakan untuk menyiasat perubahan transkriptomik akar kelapa sawit (*Elaeis guineensis* Jacq.) (Deli x Yangambi) *in vitro* yang berumur dua bulan selepas diinokulasi dengan *B. sphaericus* UPMB10 selama 120 jam. Kajian ini bermula dengan penjanaan 1824 penanda jujukan terekspres (EST) daripada akar kelapa sawit. Sejumlah 1173 gen unik yang tentatif (TUG) telah dikelompokkan daripada 1566 EST dengan jujukan yang boleh dibaca. Walau bagaimanapun, hanya 984 TUG menunjukkan pepadanan bermakna (nilai E kurang daripada  $10^{-5}$ ) dengan jujukan dalam pangkalan data protein yang tidak berulang dan TUG dibahagikan



kepada 13 kumpulan berdasarkan fungsi putatif. Keputusan mikroatur yang seterusnya menunjukkan bahawa 151 dan 125 transkrip mengalami pengawalaturan naik dan turun yang signifikan dalam akar kelapa sawit yang diinokulasi dengan *B. sphaericus* UPMB10. Walaupun transkrip yang terlibat dalam penghasilan protein telah dikawalatur naik dan paras ekspresi gen-gen yang bereaksi terhadap auksin telah berubah dalam kelapa sawit yang diinokulasi dengan *B. sphaericus* UPMB10, tiada keputusan yang muktamad untuk menyokong kehadiran auksin yang dirembeskan oleh *B. sphaericus* UPMB10 dalam medium. Walaupun *B. sphaericus* UPMB10 mempunyai kebolehan untuk mengikat nitrogen daripada udara, tempoh 120 jam mungkin tidak cukup untuk *B. sphaericus* UPMB10 mewujudkan pengikatan nitrogen yang berkesan untuk membebaskan kelapa sawit daripada kekurangan nitrogen. Interaksi di antara tumbuhan dan bakteria mungkin mengaktifkan pertahanan tumbuhan yang menyebabkan pengawalaturan naik transkrip yang berkaitan dengan penghasilan hidrogen peroksida dalam kelapa sawit. Pengesahan keputusan mikroatur dengan RT-PCR masa-nyata (real time RT-PCR) menunjukkan bahawa corak ekspresi sembilan daripada 11 calon gen (81.8%) adalah konsisten. Kesimpulannya, kajian ini telah memberikan satu pemahaman yang ringkas terhadap perubahan transkriptomik di kelapa sawit yang diinokulasi dengan *B. sphaericus* UPMB10.

## ACKNOWLEDGEMENTS

I would like to express my deepest gratitude to Assoc. Prof. Dr. Ho Chai Ling and Prof. Zulkifli Hj. Shamsuddin for their advice and guidance; Dr. Tan Siang Hee and Ng Wai Har for cDNA library and cDNA clones of root from oil palm; Dr. Harikrishna Kulaveerasingam, Dr. Meilina Ong Abdullah, Lee Yang Ping, Kwan Yen Yen, and Tee Jin Meng for cDNA clones of vegetative meristem and zygotic embryo; Dr. Maheran Abu Bakar and Aw Khoo Teng for preparation of *in vitro* oil palm seedling; Irni Suhayu Bt. Sapian for microarray scanning and Dr. Premalatha Pakirisamy and all laboratory members especially Siti Habshah, Tee Sue Sean, Teo Chin Jit, Teo Swee Sen, Teoh Seddon and Yong Sock Hwa for their assistance and support throughout this study. This work was financially supported by grant IRPA 01-04-03-T0045-TC2 from the Ministry of Science, Technology and Innovation of Malaysia.



I certify that an Examination Committee has met on 11 June 2008 to conduct the final examination of Lim Kok Ang on his Master of Science thesis entitled "Analysis of Transcriptomic Changes in Oil Palm (*Elaeis guineensis* Jacq.) Root upon Inoculation with *Bacillus sphaericus* UPMB10 Using cDNA Microarray" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the Master of Science.

Members of the Examination Committee were as follows:

**Raha Abdul Rahim, PhD**

Professor  
Faculty of Biotechnology and Biomolecular Science  
Universiti Putra Malaysia  
(Chairman)

**Parameswari a/p Namasivayam, PhD**

Lecturer  
Faculty of Biotechnology and Biomolecular Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Wong Mui Yun, PhD**

Lecturer  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Mohd. Nazalan Mohd. Najimudin, PhD**

Professor  
School of Biological Science  
Universiti Sains Malaysia  
Malaysia  
(External Examiner)

---

**HASANAH MOHD GHAZALI, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date :





This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee are as follows:

**Ho Chai Ling, PhD**

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Zulkifli Hj. Shamsuddin, PhD**

Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Member)

---

**AINI IDERIS, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date : 14 August 2008



## **DECLARATION**

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

**LIM KOK ANG**

Date: 7 July 2008



## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	<b>ii</b>
<b>ABSTRAK</b>	<b>iv</b>
<b>ACKNOWLEDEMENTS</b>	<b>vi</b>
<b>APPROVAL</b>	<b>vii</b>
<b>DECLARATION</b>	<b>ix</b>
<b>LIST OF TABLES</b>	<b>xiii</b>
<b>LIST OF FIGURES</b>	<b>xiv</b>
<b>LIST OF ABBREVIATIONS</b>	<b>xv</b>
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	
2.1 Oil Palm	4
2.1.1 Botany of Oil Palm	4
2.1.2 Root of Oil Palm	5
2.1.3 History of Oil Palm in Malaysia	8
2.1.4 Application of Fertilizer in Plantation of Oil Palm	9
2.2 Plant-Microbe Interaction	11
2.2.1 Plant-Plant Growth Promoting Bacteria (PGPB) Interactions	12
2.2.2 Biological Nitrogen Fixation	16
2.2.3 Production of Plant Growth Promoting Substances	19
2.2.4 <i>Bacillus sphaericus</i>	21
2.3 Functional Genomics	22
2.3.1 Transcriptomics	23
2.3.2 Expressed Sequence Tags Sequencing	24
2.3.3 Microarray	26
2.3.4 Real Time Reverse Transcription-PCR (Real Time RT-PCR)	30
<b>3 MATERIALS AND METHODS</b>	
3.1 Sample Materials	34
3.2 Generation of Expressed Sequence Tags (ESTs)	34
3.2.1 Preparation of Host Bacteria	34
3.2.2 Titering the Amplified cDNA Library of Root from Oil Palm and Blue-White Selection with IPTG and X-Gal	35
3.2.3 Mass Excision of the Amplified cDNA Library of Root from Oil Palm	36



3.2.4	Transformation of <i>Escherichia coli</i> SOLR with Excised Amplified cDNA Library from The Root of Oil Palm	37
3.2.5	Plasmid Isolation by Using Perfectprep® Plasmid 96 Vac, Direct Bind	37
3.2.6	DNA Sequencing of cDNA Clones	39
3.2.7	Bioinformatic Analysis of ESTs	39
3.3	Fabrication of cDNA microarray slide	40
3.3.1	PCR Amplification of cDNA Clones	40
3.3.2	Purification of PCR Products and Gel Electrophoresis	41
3.3.3	Preparation of Lucidia Universal ScoreCard Controls	42
3.3.4	Arraying of cDNA Probes	42
3.4	Inoculation of <i>In-vitro</i> Oil Palm Plantlets by <i>Bacillus sphaericus</i> UPMB10	43
3.4.1	Preparation of <i>B. sphaericus</i> UPMB10 Culture	43
3.4.2	Inoculation of Oil Palm Plantlets (Deli x Yangambi) with <i>B. sphaericus</i> UPMB10 Culture and Sample Harvesting	43
3.5	RNA Extraction from the Root of Oil Palm Using Modified CTAB Method	44
3.6	Microarray Hybridization	46
3.6.1	Amplification of RNA by Using MessageAmp™ II aRNA Amplification Kit	46
3.6.2	Preparation of Fluorescently Labelled cDNA from aRNA by Using Cyscribe First-strand cDNA Labelling Kit (Amersham Biosciences)	48
3.6.3	Purification of Labelled cDNA with CyScribe GFX Purification Kit	49
3.6.4	cDNA Microarray Hybridization	49
3.6.5	Microarray Data Analysis	50
3.7	Verification of Microarray Result Using Real Time RT-PCR	51
3.7.1	Primer Design	51
3.7.2	Treatment of RNA Sample with DNaseI	52
3.7.3	Synthesis of First Strand cDNA by using StrataScript® QPCR cDNA Synthesis Kit	53
3.7.4	Real Time RT-PCR by Using Brilliant® SYBR® Green QPCR Master Mix	54
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	
4.1	Generation of ESTs from the Roots of Oil Palm	55
4.2	Fabrication of cDNA Microarray Slide	66
4.3	RNA Extraction	68
4.4	Microarray Analysis	73
4.5	Verification of Microarray Result Using Real Time RT-PCR	85



<b>5</b>	<b>CONCLUSIONS</b>	89
	<b>REFERENCES</b>	91
	<b>APPENDICES</b>	102
	<b>BIODATA OF STUDENT</b>	139



## LIST OF TABLES

Table		Page
4.1	Characteristics of ESTs from the root of oil palm	56
4.2	Redundancy of ESTs in different cDNA libraries of oil palm	58
4.3	The TUCs with EST redundancy ranged from 6 to 28.	63
4.4	Quality and yield of RNA extracted with modified CTAB method and amplified RNA (aRNA)	72
4.5	The fold changes of auxin responsive genes in <i>B. sphaericus</i> UPMB10-inoculated oil palm	79
4.6	The fold changes of transcripts involved in the nitrogen metabolism in <i>B. sphaericus</i> UPMB10-inoculated oil palm	82
4.7	The fold changes of transcripts involved in the plant defense in <i>B. sphaericus</i> UPMB10-inoculated oil palm	84
4.8	The fold changes of the genes obtained by microarray and real time RT-PCR from the oil palm roots harvested at 5 days after inoculation with <i>B. sphaericus</i> UPMB10	87



## LIST OF FIGURES

Figure		Page
2.1	Root system of seedling 36 days after germination (a), and 3-month-old oil palm (b)	7
2.2	Detection chemistry of real time RT-PCR using sequence specific probes	31
4.1	Functional classification of ESTs from the root of oil palm	59
4.2	RNA extracted from root of oil palm by using CTAB extraction buffer modified by Wang <i>et al.</i> (2005b) were separated on 1 % denaturing formaldehyde gel.	71
4.3	Classification of the transcripts up-regulated in oil palm inoculated with <i>B. sphaericus</i> UPMB10	76
4.4	Classification of the transcripts down-regulated in oil palm inoculated with <i>B. sphaericus</i> UPMB10	77



## LIST OF ABBREVIATIONS

$\Delta$	Delta
®	Registered trademark
1 x	1 time
10 x	10 times
<sup>15</sup> N	Nitrogen-15
24:1	Ratio 24 to 1
25:24:1	Ratio 25 to 24 to 1
6 x	6 times
A <sub>260/230</sub>	Absorbance at wavelength of 260 nm over 230 nm
A <sub>260/280</sub>	Absorbance at wavelength of 260 nm over 280 nm
ADP	Adenine diphosphate
aRNA	Amplified RNA
ATP	Adenosine triphosphate
BNF	Biological nitrogen fixation
bp	Base pair
CaCl <sub>2</sub>	Calcium chloride
CAP3	Contig Assembly Program 3
cDNA	Complementary DNA
cfu	Colonies forming unit
CsCl	Cesium chloride
Ct	Threshold cycle
CTAB	Cetyl trimethyl ammonium bromide
CTP	Cytidine triphosphate
dbEST	Database of EST
dCTP	2' deoxycytidine 5' -triphosphate
DEPC	Diethyl pyrocarbonate
DMSO	Dimethylsulfoxide
DNaseI	Deoxyribonuclease I





dNTP	2'-deoxyribonucleoside triphosphate
DRAG	dinitrogenase reductase-activating glycohydrolase
DRAT	Dinitrogenase reductase ADP-ribosyltransferase
DTT	Dithiothreitol
E	Efficiency
e <sup>-</sup>	Electron
EDTA	Ethylenediamne tetraacetic acid
EST	Expressed sequence tag
FDR	False discovery rate
Fe	Iron
FELCRA	Federal Land Consolidation and Rehabilitation Authority
FELDA	Federal Land Development Authority
<i>g</i>	Relative centrifugal force ( <i>rcf</i> )
GTP	Guanosine triphosphate
H <sup>+</sup>	Proton
H <sub>2</sub>	Hydrogen
HCl	Hydrochloric acid
HEPES	<i>N</i> -(2-hydroxyethyl)piperazine- <i>N'</i> -(2-ethanesulfonic acid)
IAA	Indole-3-acetic acid
IPTG	Isopropylthio-β-D-galactoside
ISR	Induced systemic resistance
KCl	Potassium chloride
KEGG	Kyoto Encyclopedia of Genes and Genomes
kPa	KiloPascal
LB	Luria-Bertani
LiCl	Lithium chloride
M	Molar
MgCl <sub>2</sub>	Magnesium chloride
MgSO <sub>4</sub>	Magnesium sulfate
mM	Milimolar
Mo	Molybdenum



mRNA	Messenger RNA
MS	Murashige and Skoog
N	Nitrogen
N <sub>2</sub>	Nitrogen
NAA	Naphthalene acetic acid
NaCl	Sodium chloride
NAD	Nicotinamide adenine dinucleotide
NaOH	Sodium hydroxide
NCBI	National Center for Biotechnology Information
Ndfa	Nitrogen derived from atmosphere
ng	Nanogram
NH <sub>3</sub>	Ammonia
NH <sub>4</sub> <sup>+</sup>	Ammonium
nM	Nanomolar
OD <sub>600</sub>	Optical density at wavelength of 600nm
PBS	Phosphate-buffered saline
PCR	Polymerase Chain Reaction
Pfu	Plaques forming unit
PGPB	Plant growth promoting bacteria
PGPR	Plant growth promoting rhizobacteria
P <sub>i</sub>	Phosphate
PVPP	Polyvinylpyrrolidone
Real Time RT-PCR	Real time reverse transcription-PCR
RI <sub>1-5</sub>	Primary roots emitted from a 1 to 5-month-old oil palm
RI <sub>5-12</sub>	Primary roots emitted from a 5 to 12-month-old oil palm
RISDA	Rubber Industry Smallholders' Development Authority
RNA	Ribonucleic acids
RNase	Ribonuclease
ROS	Reactive oxygen species
Rpm	Revolution per minute
SAM	Significant analysis of microarray

SDS	Sodium dodecyl sulfate
sp.	species
SSC	Standard saline citrate
TAE	Tris-acetate EDTA
T <sub>m</sub>	Melting temperature
™	Trade mark
Tris	2-amino-2-hydroxymethyl-1,3-propanediol
TS	Tryptic soy
TUC	Tentative unique contig
TUG	Tentative unique gene
TUS	Tentative unique sequence
U	Unit
UTP	Uridine triphosphate
v/v	Volume per volume
w/v	Weight per volume
X-gal	5-bromo-4-chloro-3-indolyl-β-D-galactoside
β-	Beta-
δ	delta
λ	Lambda
μg	Microgram
μL	Microlitre
μM	Micromolar



# CHAPTER 1

## INTRODUCTION

In the soil surrounding the plant root, the release of root exudates has attracted the colonization of plants by bacteria. The bacteria that are beneficial to plants are collectively known as plant growth-promoting bacteria (PGPB). The major difference between the PGPB and *Rhizobium* is that PGPB do not form a distinct symbiotic relationship with the host plant. Generally, PGPB might establish an endophytic, rhizospheric or associative relationship with the host plant and this depends on the combination of PGPB and host plant. Various mechanisms have been employed by PGPB to improve plant growth, including biological nitrogen fixation, production of plant growth promoting substances which enhance the root development, stimulation of plant defense to suppress the growth of pathogen or combination of modes of mechanisms (Vessey, 2003).

Oil palm (*Elaeis guineensis*) is one of the most important plantation crops in Malaysia and a major source of edible oil in the world. The planting of oil palm requires a high input of chemical fertilizer, thus posing severe threats to the environment such as emission of greenhouse gases, nitrate pollution and acidification of soil. Amir *et al.* (2002) reported oil palm inoculated with a local isolate of PGPB, *Bacillus sphaericus* UPMB10 showed an enhanced growth. Besides reducing the application of chemical fertilizer, inoculation of oil palm with *B. sphaericus* UPMB10 could reduce the production cost, leading toward a more sustainable management.



Amir *et al.* (2002) attributed the enhanced oil palm growth to the ability of *B. sphaericus* to fix atmospheric nitrogen and synthesize plant growth promoting substances such as auxin. However, the detailed understanding of interaction between oil palm and *B. sphaericus* UPMB10 is still lacking. Although PGPB are widely studied, the understanding of their interaction with PGPB and plant is not as good as that between *Rhizobium* and legume. This might be due to the highly diversified PGPB and the lack of specific host plant. A comprehensive transcriptomic study can reveal how the interaction is initiated and developed between oil palm and *B. sphaericus* UPMB10, and can explain mechanism(s) employed by PGPB to promote plant growth. To accelerate the understanding of the interaction between *B. sphaericus* UPMB10 and oil palm, cDNA microarray, a high throughput approach has been used to elucidate the transcriptomic event of oil palm upon inoculation with *B. sphaericus* UPMB10.

Since oil palm is not a model plant and the cDNA sequences in the public domain are limited, expressed sequence tags (ESTs) were generated and used as cDNA probes for cDNA microarray. In this study, the RNA derived from the root of *B. sphaericus* UPMB10-inoculated oil palms and reference controls (without the inoculation with *B. sphaericus* UPMB10) were labeled with fluorescent dyes and co-hybridized to oil palm microarray. Although microarray is a high throughput approach, it is very expensive to use and requires complicated statistic analysis of data. In addition to that, post microarray verification such as real time PCR and Northern analysis is necessary.

The objectives of this study were:

1. To generate ESTs from the oil palm root cDNA library to be used as cDNA probes for cDNA microarray.
2. To study the transcriptomic changes in the oil palm root upon inoculation of *B. sphaericus* UPMB10 by using cDNA microarray approach.
3. To verify the microarray results by using real time PCR.

## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Oil Palm

##### 2.1.1 Botany of Oil Palm

The oil palm, *Elaeis guineensis* Jacq is classified together with *Cocos* (the coconut) and other genera under the subfamily Cocosoidae, in the family Aracaceae (formerly known as Palmae) (Corley and Tinker, 2003). The botanical name of *E. guineensis* was given by Jacquin. The genus name *Elaeis* is derived from the Greek word *elaion*, meaning oil while the specific name *guineensis* shows its origin in the Guinea Coast, in West Africa. The genus *Elaeis* consists of three species: *E. guineensis* (African oil palm), *E. oleifera* (American oil palm) and *E. odora* (formerly known as *Barcella odora*) (Corley and Tinker, 2003). *E. guineensis* is found in the tropics, within  $\pm 10^\circ$  latitude of the equator, in Africa, South East Asia and South and Central America.

*E. guineensis* has a solitary columnar stem with the crown consisting of large pinnate leave (Corley and Tinker, 2003). At the base of the stem, there is a prominent bole (bulb) from where the primary roots emerge. *E. guineensis* is normally monoecious, bearing male or female flower (Corley and Tinker, 2003). The fruit is a drupe which consists of a thin epicarp, an oily mesocarp and a nut or seed and generally borne on a large, compact bunch. The seed generally contains one to three embryos surrounding

by a large endosperm (kernel) and hard stone endocarp (shell). The oil palm root will be described in detail in the next section (Corley and Tinker, 2003).

The fruits of oil palm exhibit great variation in either the fruit type or form and these have been used for the classification of the varieties. The oil palm fruit can be divided into two types, *nigrescens* (purple) and *virescens* (green) on the basis of the external color. The thickness of the shell distinguishes the oil palm fruits into three forms: *dura* which produces fruits with a thick shell, *psifera*, which is without a shell in its rare fruits; *tenera* which is a hybrid of *dura* and *psifera* and produces fruits with an intermediate shell. However, Corley and Tinker (2003) indicated that the term variety is inappropriate for *tenera* and other forms since the materials are heterogeneous for most of the characters other than shell thickness.

### **2.1.2 Root of Oil Palm**

The root systems of terrestrial plant play two important primary roles: acquisition of resources especially water and dissolved ions from the soil and anchorage (Fitter, 1991). Other functions of the root system such as storage, synthesis of plant growth regulator and propagation are classified as secondary roles (Fitter, 1991). Root systems have been grouped into three categories based on the root diameters and the development of root hairs: graminoid, magnolioid and intermediate root systems (Ingrouille and Eddie, 2006). Graminoid root systems consist of profuse root hairs and



have very large absorptive area to root volume ratio whereas magnolioid root systems have roots with large diameter and fewer root hairs (Ingrouille and Eddie, 2006).

The oil palm has a fibrous root system. In the juvenile phase of oil palm, six types of root are found: radicle, primary roots emitted from a 1 to 5-month-old oil palm (RI<sub>1-5</sub>), primary roots emitted from 5 to 12-month-old oil palm (RI<sub>5-12</sub>), lateral long roots, lateral medium roots and lateral short roots (Figure 2.1) (Jourdan and Rey, 1997).

As the oil palm seed germinates, the radicle emerges and grows continuously in a slightly undulating pattern (Jourdan and Rey, 1997). The roots located at the base of the radicle are known as lateral long root (usually over 10 cm long) and emit very soon after germination (Jourdan *et al.*, 1995). The lateral medium roots that are less than 10 cm in length are distributed over long root and also the entire branched zone of the radicle. They bear the lateral short roots which do not exceed 1.5 cm in length and are non-branching in contrast to the lateral long and medium roots (Jourdan *et al.*, 1995).

One month after germination, the first adventitious primary root (RI<sub>1-5</sub>) emits and grows in vertical, downward direction (Jourdan and Rey, 1997). Similar to the radicle, the primary roots have the branched structure. The RI<sub>1-5</sub> and RI<sub>5-12</sub> are white when young but rapidly turn to dark brown. The lateral roots found in the RI<sub>1-5</sub> and RI<sub>5-12</sub> are similar to those of radicle but in different proportions: the proportion of long root increases with a gradual disappearance of medium and short root (Jourdan and Rey, 1997).