



**UNIVERSITI PUTRA MALAYSIA**

***PROBIOTIC POTENTIAL OF AND CONJUGATED LINOLEIC ACID  
PRODUCTION BY BACTERIA ISOLATED FROM CHICKEN AND  
RUMINANTS***

**YONG SU TING**

**FBSB 2013 36**



**PROBIOTIC POTENTIAL OF AND CONJUGATED LINOLEIC ACID  
PRODUCTION BY BACTERIA ISOLATED FROM CHICKEN AND  
RUMINANTS**

**By**

**YONG SU TING**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirement for the Degree of Master of Science**

**July 2013**



© COP YRIGHT UPM

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia





© COP YRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the degree of Master of Science

**PROBIOTIC POTENTIAL OF AND CONJUGATED LINOLEIC ACID  
PRODUCTION BY BACTERIA ISOLATED FROM CHICKEN AND  
RUMINANTS**

By

**YONG SU TING**

**July 2013**

**Chairman: Wan Zuhainis Saad, PhD**

**Faculty: Biotechnology and Biomolecular Sciences**

Conjugated linoleic acid (CLA) refers to fatty acids with 18-carbon and 2 conjugated double bonds in different positional and geometric configurations (C18:2). CLA can be normally found in ruminant meat, milk, cheese, dairy products, egg yolk and chicken. Dietary CLA intake by human is too low to exhibit health benefits such as anti-cancer, anti-inflammatory, anti-atherosclerosis, anti-obesity and modulation of immune system. Furthermore, the purity of CLA isomers is crucial for human health but the commercially available CLA was mostly produced by chemical synthesis which leads to the production of mix isomers of CLA. This can be achieved through bioproduction of CLA by bacteria and the increase of consumption of CLA enriched products by human. Therefore, the objectives of this study were to investigate the ability of bacteria isolated from ruminants, chicken, human milk and infant feces to produce CLA and to evaluate their probiotic characteristics and factors that affect the CLA production. In this study, 120 isolates were tested for the presence of catalase and Gram staining. A total of 69 isolates with catalase negative, Gram positive and

grown on MRS medium under anaerobic condition were screened for CLA production. From the screening results, 18 isolates which isolated from cattle, deer and chicken have the ability to produce CLA from lactic acid (LA). The four highest CLA-producing bacteria were isolated from chicken and cattle. They were identified as *Lactobacillus salivarius* strain P2, *Lactobacillus agilis* strain P3, *Enterococcus faecium* strain P1 and *Streptococcus equinus* strain C3 based on molecular method and produced CLA concentration of 21.97, 31.08, 23.35 and 10.23 µg/ml, respectively. *Lactobacillus salivarius* strain P2, *L. agilis* strain P3, *E. faecium* strain P1 and *S. equinus* strain C3 consist of 60.65%, 66.90%, 49.77% and 52.01% of *cis*-9, *trans*-11 (*c9*, *t11*) CLA isomer and 39.35%, 33.10%, 50.23% and 47.99% of *trans*-10, *cis*-12 (*t10*, *c12*) CLA isomer, respectively. Reference strain, *Lactobacillus reuteri* ATCC 55739 produced CLA concentration of 122.45 µg/ml which consist of 89.24% of *c9*, *t11* CLA isomer and 10.76% of *t10*, *c12* CLA isomer. All bacteria tested were able to tolerate 0.3 % bile salts and pH 2.5 acidic condition. As compared to *L. reuteri* ATCC 55739, *L. agilis* strain P3 and *L. salivarius* strain P2 showed better acidic tolerance, resistant to two types of antibiotics tested, higher antimicrobial activity and ability to produce higher amount of lactic acid. All bacterial strains showed no bacteriocin production. *Streptococcus equinus* strain C3 was resistant to four types of antibiotic tested and *E. faecium* strain P1 was hemolytic positive bacteria. These characteristics have made these 2 strains not suitable as probiotic bacteria. The most efficient CLA production were obtained by 20% cell density in citrate buffer, pH 5.5 containing 1.0 mg/ml LA which were incubated at 30°C for 48 h with shaking at 120 rpm under aerobic condition. Under optimal reaction conditions, *L. reuteri* ATCC 55739 produced the highest concentration of CLA, 280.75 µg/ml reaction mixture, followed by *L. agilis* strain P3 with 109.78 µg/ml

reaction mixture and *L. salivarius* strain P2 with 84.58 µg/ml reaction mixture. The CLA isomers formed were majority *c9, t11* CLA and minority *t10, c12* CLA. Taking into account of the probiotic effect of bacteria, *L. salivarius* strain P2 and *L. agilis* strain P3 are more suitable to be probiotic for animals and to produce CLA extracellular. The findings in this study prompt further studies to be carried out to investigate the ability of bacteria to produce CLA and their probiotic effects in animal model.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**POTENSI PROBIOTIK DAN PENGHASILAN ASID LINOLEIK  
TERKONJUGAT OLEH BAKTERIA YANG DIPENCILKAN DARIPADA  
AYAM DAN RUMINAN**

Oleh

**YONG SU TING**

**Julai 2013**

**Pengerusi: Wan Zuhainis Saad, PhD**

**Fakulti: Bioteknologi dan Sains Biomolekul**

Asid linoleik terkonjugat (CLA) merujuk kepada asid lemak dengan 18 karbon dan 2 ikatan ganda dua terkonjugat dalam konfigurasi kedudukan dan geometri yang berbeza (C18:2). CLA biasanya boleh ditemui dalam daging ruminan, susu, keju, produk tenusu, kuning telur dan ayam. Pengambilan CLA dalam diet oleh manusia adalah terlalu rendah untuk menunjukkan manfaat kesihatan seperti anti-kanser, anti-radang, anti-aterosklerosis, anti-obesiti dan modulasi sistem imun. Tambahan pula, ketulenan CLA isomer adalah penting untuk kesihatan manusia tetapi kebanyakan CLA yang didapati secara komersial adalah dihasilkan daripada sintesis kimia di mana sintesis ini membawa kepada penghasilan isomer campuran CLA. Ini boleh dicapai melalui penghasilan biologi CLA oleh bakteria dan peningkatan pemakanan produk yang diperkaya dengan CLA oleh manusia. Oleh itu, objektif kajian ini adalah untuk menyelidik keupayaan bakteria yang dipencilkan daripada ruminan, ayam, susu manusia dan najis bayi untuk menghasilkan CLA dan untuk menilai ciri-ciri probiotik bakteria dan faktor-faktor yang mempengaruhi pengeluaran CLA.

Dalam kajian ini, 120 isolat telah diuji untuk kehadiran katalase dan pewarnaan Gram. Sebanyak 69 isolat dengan katalase negatif, Gram positif dan tumbuh atas medium MRS di bawah keadaan anaerobik telah disaring untuk penghasilan CLA. Daripada keputusan penyaringan, 18 isolat yang dipencilkan daripada lembu, rusa dan ayam mempunyai keupayaan untuk menghasilkan CLA daripada asid laktik (LA). Empat bakteria yang mempunyai penghasilan CLA yang tertinggi adalah dipencilkan daripada ayam dan lembu. Bakteria telah dikenalpasti sebagai strain P2 *Lactobacillus salivarius*, strain P3 *Lactobacillus agilis*, strain P1 *Enterococcus faecium* dan strain C3 *Streptococcus equinus* berdasarkan cara molekul dan menghasilkan kepekatan CLA dengan 21.97, 31.08, 23.35 dan 10.23 µg/ml, masing-masing. Strain P2 *Lactobacillus salivarius*, strain P3 *L. agilis*, strain P1 *E. faecium* dan strain C3 *S. equines* terdiri daripada 60.65%, 66.90%, 49.77% dan 52.01% isomer CLA *cis*-9, *trans*-11 (*c*9, *t*11) dan 39.35%, 33.10%, 50.23% dan 47.99% isomer CLA *trans*-10, *cis*-12 (*t*10, *c*12), masing-masing. Strain rujukan, *Lactobacillus reuteri* ATCC 55739 menghasilkan kepekatan CLA dengan 122.45 µg/ml yang terdiri daripada 89.24% isomer CLA *c*9, *t*11 dan 10.76% isomer CLA *t*10, *c*12. Semua bakteria yang diuji boleh toleransi kepada 0.3% garam hempedu dan keadaan asid pH 2.5. Jika berbanding dengan *L. reuteri* ATCC 55739, strain P3 *L. agilis* dan strain P2 *L. salivarius* menunjukkan toleransi asid yang lebih baik pada pH 2.5, rintangan terhadap dua jenis antibiotic yang diuji, aktiviti antimikrob yang lebih tinggi dan keupayaan untuk menghasilkan jumlah asid laktik yang lebih tinggi. Semua strain bakteria menunjukkan tiada penghasilan bakteriosin. Strain C3 *Streptococcus equines* mempunyai rintangan terhadap empat jenis antibiotik yang diuji dan strain P1 *E. faecium* menunjukkan hemolitik positif. Ciri-ciri ini telah menjadikan 2 strain ini tidak sesuai sebagai bakteria probiotik. Penghasilan CLA

yang paling efisien diperolehi dengan 20% kepadatan sel dalam penimbal sitrat, pH 5.5 yang mengandungi 1.0 mg/ml LA yang dieram pada 30°C selama 48 jam dengan 120 rpm di bawah keadaan aerobik. Di bawah keadaan reaksi yang optimum, *L. reuteri* ATCC 55739 menghasilkan kepekatan CLA yang tertinggi, 280.75 µg/ml campuran, diikuti oleh strain P3 *L. agilis* dengan 109.78 µg/ml campuran dan strain P2 *L. salivarius* dengan 84.58 µg/ml campuran. Isomer CLA yang terbentuk majoriti adalah *c9*, *t11* CLA dan minoriti adalah *t10*, *c12* CLA. Jika kesan probiotik diambil kira, strain P2 *Lactobacillus salivarius* dan strain P3 *L. agilis* adalah lebih sesuai untuk dijadikan sebagai probiotik untuk haiwan dan menghasilkan CLA ekstrasellular. Penemuan dalam kajian ini mewajarkan kajian lebih lanjut dijalankan untuk menyiasat keupayaan bakteria menghasilkan CLA dan kesan probiotik dalam model haiwan.

## ACKNOWLEDGEMENTS

I would like to express my deepest gratitude to my main supervisor Dr. Wan Zuhainis Saad for her constant encouragement, patient and support during the course of the study. She did not only devote her invaluable time to teach, advice and support my academic activities but also always had solutions for any problem. I would like to express my sincere thanks to my supervisory committee, Assoc. Prof. Dr. Sieo Chin Chin for her valuable advice and feel very thankful to work with her. Special thanks to Prof. Dr. Ho Yin Wan who provided me the great opportunity to carry out the research in her laboratory at Laboratory of Vaccines and Immunotherapeutics, Institute of Bioscience (IBS), Universiti Putra Malaysia. Thanks for her wealth of knowledge, critical comments and guidance during the planning and progress of the study.

Sincere thanks to my friend, Tan Hui Yin for her knowledge, thinking and encouragement throughout the study. I am thankful for her helpful advice in the preparation of manuscripts and statistical analysis. Thanks to her who listens when I am angry and laugh with or at me whenever possible. Thanks for being there!

I would like to thank to my friend, Mahdi for his valuable help and assistance in the analysis of samples for fatty acids profiles. His contributions and technical advice on gas chromatography really helped me a lot throughout the hardest time during my study.

To all of the supporting staff and lovely friends at the IBS laboratory, thank you very much for their assistance, friendships and parties. They are Madam Haw, Encik Khairul, Mr Nagayah, Pornpan, Sam, Azim, Ihsan, Gee Leng, Chuan Loo, Pui Wah, Xiao Dan, Xiao Jing, Wei Li, Wanqin, Ainn, Zira, Khomala, Parisa, Mohammad and Ehsan. It was my great pleasure knowing all of them. Sharing of emotions and experiences with them was the most enjoyable and valuable time.

Special thanks to my housemates and friends, Shi Wei, Poh Tee, Ching Mun, Tse Peng, Pei Shen, Eileen, Chia Yean, Ming Yuen, and Seow Ching for their wonderful friendship and encouragement to me from time to time. I am most grateful to my parents and family members for their constant and unconditional encouragement.

Finally, I would like to acknowledge Universiti Putra Malaysia for granting me the Graduate Research Fellowship. I also would like to take this opportunity to thank everyone that has contributed to my personal and educational development.

I certify that a Thesis Examination Committee has met on 23 July 2013 to conduct the final examination of Yong Su Ting on her thesis entitled “Probiotic Potential of and Conjugated Linoleic Acid Production by Bacteria Isolated from Chicken and Ruminants” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Janna Ong binti Abdullah, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Shuhaimi bin Mustafa, PhD**

Professor  
Halal Products Research Institute  
Universiti Putra Malaysia  
(Internal Examiner)

**Syahida binti Ahmad, PhD**

Senior lecturer  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Kalavathy Ramasamy, PhD**

Associate Professor  
Faculty of Pharmacy  
Universiti Teknologi Mara  
(External Examiner)

---

**NORITAH OMAR, PhD**

Associate Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 19 September 2013

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Wan Zuhainis Saad, PhD**

Senior lecturer  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Ho Yin Wan, PhD**

Professor  
Institute of Bioscience  
Universiti Putra Malaysia  
(Member)

**Sieo Chin Chin, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

---

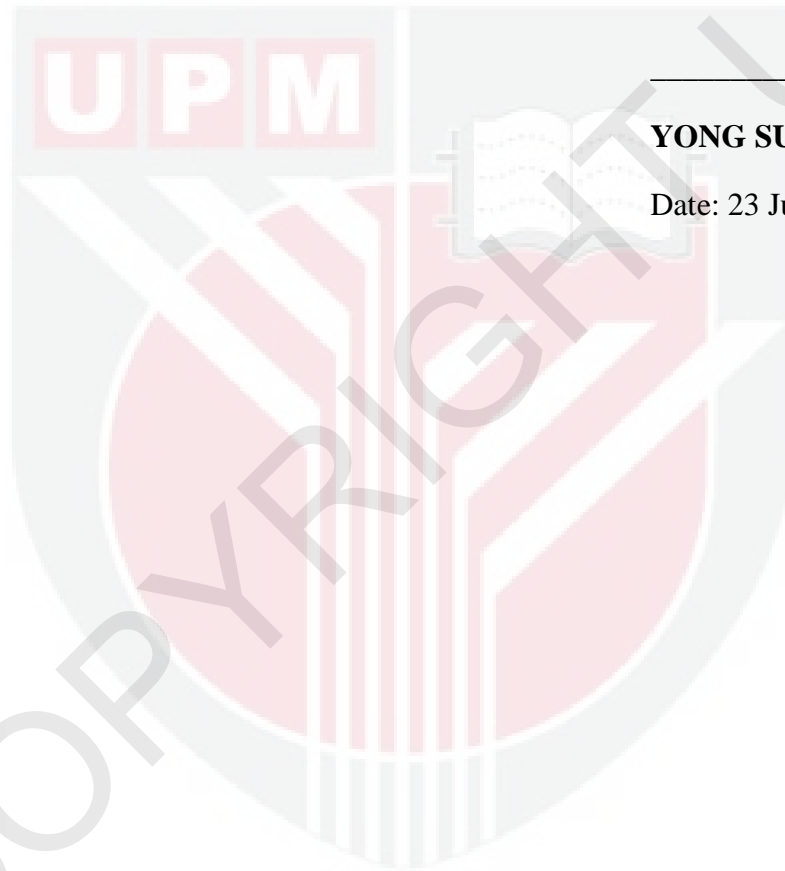
**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or other institutions.



---

**YONG SU TING**

Date: 23 July 2013



## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	ii
<b>ABSTRAK</b>	v
<b>ACKNOWLEDGEMENTS</b>	viii
<b>APPROVAL</b>	x
<b>DECLARATION</b>	xii
<b>LIST OF TABLES</b>	xvi
<b>LIST OF FIGURES</b>	xvii
<b>LIST OF ABBREVIATIONS</b>	xx
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	<b>5</b>
2.1 Conjugated linoleic acid (CLA) and its functional properties	5
2.1.1 Chemical structure of CLA	7
2.1.2 Occurrence of CLA in ruminants and nonruminants	8
2.1.3 Occurrence of CLA in human	9
2.1.4 Occurrence of CLA in foods	11
2.1.5 Anticarcinogenic properties of CLA	13
2.1.6 Antiatherosclerotic properties of CLA	15
2.1.7 Modulation of immune system by CLA	17
2.1.8 Antiobesity properties of CLA	18
2.1.9 Effects in insulin sensitivity of human	20
2.2 Synthesis of CLA	20
2.2.1 Formation of CLA by chemical method	20
2.2.2 Formation of CLA in the rumen	21
2.2.3 Formation of CLA in milk and animal tissues	24
2.2.4 Formation of CLA by bacteria	24
2.3 Probiotic lactic acid bacteria	25
2.3.1 Probiotic bacteria in the improvement of animal health	26
2.3.2 CLA producing probiotic lactic acid bacteria	27
2.4 Analysis of CLA	28
2.4.1 Extraction of lipid	28
2.4.2 Quantification of CLA	28
2.4.3 Qualitative determination of CLA	29
2.4.4 Methylation procedures for CLA	29
2.4.5 Separation and identification of CLA isomers	32
<b>3 ISOLATION, SCREENING AND IDENTIFICATION OF CLA PRODUCING BACTERIA</b>	<b>35</b>
3.1 Introduction	35
3.2 Materials and methods	36

3.2.1	Sources of sample for isolation of lactic acid bacteria	36
3.2.2	Anaerobic technique	38
3.2.3	Preparation of media	38
3.2.4	Isolation and purification of facultative anaerobic and anaerobic bacteria	40
3.2.5	Maintenance of facultative anaerobic and anaerobic bacterial isolates	41
3.2.6	Catalase test of bacteria	41
3.2.7	Gram staining of bacteria	42
3.2.8	Screening for CLA production capabilities of bacteria	43
3.2.9	Identification of CLA producing lactic acid bacteria	46
3.2.10	Statistical analysis	48
3.3	Results and discussion	49
3.3.1	Catalase test and Gram staining of bacteria	49
3.3.2	Screening of CLA production by bacteria	49
3.3.3	Identification of CLA producing lactic acid bacteria based on morphological characteristics	54
3.3.4	Identification of CLA producing lactic acid bacteria based on molecular characteristics	59
3.4	Conclusion	64
<b>4</b>	<b>PROBIOTIC CHARACTERISTIC OF LACTIC ACID BACTERIA</b>	<b>66</b>
4.1	Introduction	66
4.2	Materials and methods	67
4.2.1	Acid tolerance test of bacteria	67
4.2.2	Bile tolerance test of bacteria	68
4.2.3	Antibiotic susceptibility test	69
4.2.4	Antibacterial activity of bacteria	70
4.2.5	Hemolytic activity of bacteria	72
4.2.6	Statistical analysis	72
4.3	Results and discussion	73
4.3.1	Acid tolerance of bacteria	73
4.3.2	Bile tolerance of bacteria	75
4.3.3	Antibiotic susceptibility test of bacteria	77
4.3.4	Antibacterial activity of bacteria	82
4.3.5	Hemolytic activity of bacteria	87
4.4	Conclusion	88
<b>5</b>	<b>EFFECT OF DIFFERENT PHYSICAL FACTORS ON THE PRODUCTION OF CLA BY BACTERIAL ISOLATES</b>	<b>89</b>
5.1	Introduction	89
5.2	Materials and methods	90
5.2.1	Preparation of washed cells	90
5.2.2	Determination of CLA production by bacterial	90

isolates under aerobic and anaerobic conditions	
5.2.3 Effect of pH on CLA production by bacterial isolates	91
5.2.4 Influence of temperature on CLA production by bacterial isolates	91
5.2.5 Effect of concentration of linoleic acid on production of CLA by bacterial isolates	92
5.2.6 Effect of cell density on CLA production by bacterial isolates	92
5.2.7 Lipid analysis	93
5.2.8 Statistical analysis	
5.3 Results and discussion	93
5.3.1 CLA production by bacterial isolates under aerobic and anaerobic conditions	93
5.3.2 CLA production by bacterial isolates at different pH	95
5.3.3 CLA production by bacterial isolates in different temperature	97
5.3.4 CLA production by bacterial isolates with different concentration of linoleic acid	98
5.3.5 CLA production by bacterial isolates with different cell density	99
5.3.6 CLA and other fatty acids profile of supernatant in optimum condition by bacterial isolates	102
5.4 Conclusion	104
<b>6 GENERAL DISCUSSION, RECOMMENDATIONS FOR FUTURE RESEARCH AND CONCLUSIONS</b>	<b>105</b>
<b>REFERENCES</b>	<b>111</b>
<b>APPENDICES</b>	<b>135</b>
<b>BIODATA OF STUDENT</b>	<b>139</b>
<b>LIST OF PUBLICATIONS</b>	<b>140</b>