

UNIVERSITI PUTRA MALAYSIA

ANALYSIS OF FLAVONOIDS AND ESSENTIAL OILS FROM CLAUSENA EXCAVATA AND THEIR MEDICINAL PROPERTIES

LIM LAY SEAN

FBSB 2004 9



ANALYSIS OF FLAVONOIDS AND ESSENTIAL OILS FROM CLAUSENA EXCAVATA AND THEIR MEDICINAL PROPERTIES

LIM LAY SEAN

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA

2004

This work is specially dedicated to

My beloved family, friends and teachers

who show me the fund of education

and as my sources of encouragement and support in the completion of this study

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

ANALYSIS OF FLAVONOIDS AND ESSENTIAL OILS FROM *CLAUSENA EXCAVATA* AND THEIR MEDICINAL PROPERTIES

By

LIM LAY SEAN

November 2004

Chairman: Associate Professor Radzali B. Muse, Ph.D.

Faculty: Biotechnology and Biomolecular Sciences

This study was conducted mainly to determine the major flavonoid compounds and the composition of essential oils predominant in Clausena excavata, and also to examine its antioxidant, antibacterial and anticancer activities. The dried yields of C. excavata's leaf crude extracts were also determined and methanol was found to be the best solvent for extracting soluble bioactive compounds from the leaves of C. excavata. Total phenolic contents were found abundantly in methanol crude extract of leaves. In the high performance liquid chromatography (HPLC) analysis, rutin and naringin were found predominant in the leaves of C. excavata, while in the gas chromatography-mass spectrometry (GC-MS) analysis, the major chemical components identified in the fruit oil were α -ocimene and terpinolene. Terpinolene was also being the main component and remarkably present in the leaf oil. Both of the fruit and leaf oils extracted with dichloromethane in the simultaneous distillation extraction (SDE) method were indicated in very low yield (less than 0.1%). Antioxidant activities of various crude extracts of Clausena excavata leaves were dependent on the amount of total phenolics present in the crude extracts. Inhibition of lipid peroxidation and free radical scavenger potential of those crude extracts closely dependent on the particular substitution pattern of free hydroxyl groups on

iii

the flavonoid skeleton. Antioxidant activity of methanol crude leaf's extract was found greater than α-tocopherol in the ferric thiocyanate (FTC) and thiobarbituric acid (TBA) methods as well. The increase in the concentration of methanol crude leaf extract from 200 to 1000 µg/ml had increased its antioxidant activity. However, methanol crude leaf's extract exhibited weak scavenging activity towards 1,1diphenyl-2-picrylhydrazyl (DPPH) free radical. As for the determination of antibacterial activity in both of the disc diffusion and microdilution methods, the growth of all tested gram-positive bacteria, especially *Micrococcus luteus* was found effectively affected by the methanol crude leaf extract. Almost all of the crude extracts were active towards Micrococcus luteus. Fruit and leaf oil exhibited their effects on all tested gram-positive bacteria and certain gram-negative bacteria but those tested bacteria strains were just weakly inhibited. Antioxidant activity of flavonoids was associated with anticancer properties. In the MTT assay, however, fruit oil showed better cytotoxic activity than that in methanol crude leaf extract against MCF-7 cells. This implied that C. excavata fruit oil might be a good source for the breast cancer treatment due to the presence of possible active anticancer agents.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

ANALISIS KE ATAS FLAVANOID DAN MINYAK PATI DARIPADA CLAUSENA EXCAVATA DAN CIRI-CIRI PERUBATANNYA

Oleh

LIM LAY SEAN

November 2004

Pengerusi: Profesor Madya Radzali B. Muse, Ph.D.

Fakulti: Bioteknologi dan Sains Biomolekul

Kaji selidik ini bertujuan untuk menentukan sebatian flavonoid yang utama dan komposisi minyak pati dalam Clausena excavata, juga untuk memeriksa aktiviti anti-oksida, anti-bakteria dan anti-kanser masing-masing. Kadar kering hasil ekstrak mentah daun C. excavata juga ditentukan dan methanol telah dikenal pasti sebagai pelarut terbaik untuk mengekstrak sebatian bio-aktif yang larut air daripada daun C. excavata. Jumlah kandungan fenolik yang tinggi didapati dalam ekstrak mentah metanol. Dalam analisis HPLC, rutin dan naringin didapati hadir dalam kandungan yang tinggi dalam daun C. excavata, sementara dalam GC-MS analisis, komponen kimia utama yang dijumpai dalam minyak pati buah ialah α-ocimene dan terpinolene. Terpinolene juga muncul sebagai komponen utama dalam minyak pati daun dengan kandungan yang cukup menakjubkan. Penghasilan minyak pati daripada buah dan daun yang diekstrak dengan diklorometana adalah sangat sedikit (kurang daripada 0.1%). Aktiviti-aktiviti anti-oksida pelbagai ekstrak mentah daripada daun C. excavata adalah bergantung kepada jumlah sebatian fenolik yang hadir di dalam sesuatu esktrak itu. Potensi ekstrak-ekstrak mentah itu sebagai penghalang bagi pengoksidaan lipid dan penderma atom hydrogen kepada radikal bebas adalah sangat bergantung kepada corak penggantian yang khusus kumpulan hidroksil bebas dalam

V

rangka struktur flavonoid. Dalam kedua-dua ujian FTC dan TBA, aktiviti anti-oksida ekstrak mentah metanol daripada daun didapati lebih berkesan daripada α-tokoferol. Peningkatan kepekatan ekstrak mentah metanol daripada daun dari 200 kepada 1000 µg/ml telah pun meningkatkan aktiviti anti-oksida. Akan tetapi, ekstrak mentah metanol daripada daun kurang berkesan dalam tindak balas terhadap radikal bebas 1,1-diphenyl-2-picrylhydrazyl (DPPH). Dalam penentuan aktiviti anti-bakteria, kaedah penyebaran dengan penggunaan cakera kertas dan kaedah mikro-pencairan medium telah diaplikasikan. Ekstrak mentah metanol daripada daun menunjukkan keberkesanannya untuk merencat pertumbuhan kesemua gram-positif bakteria yang dikaji, terutamanya terhadap *Micrococcus luteus*. Hampir kesemua ekstrak mentah adalah aktif terhadap Micrococcus luteus. Minyak pati yang diekstrak daripada kedua-dua buah dan daun menunjukkan aktiviti perencatan ke atas semua grampositif bakteria dan juga sesetengah gram-negatif bakteria, akan tetapi, kesan perencatan tersebut agak lemah. Aktiviti anti-oksida flavonoid berhubung-kait dengan anti-kanser. Dalam ujian MTT, walau bagaimanapun, minyak pati daripada buah memaparkan kesan sitotoksik yang lebih kuat terhadap sel MCF-7 jika dibandingkan dengan ekstrak mentah metanol daripada daun. Ini menunjukkan bahawa minyak pati daripada buah mempunyai potensi sebagai sumber yang baik untuk rawatan kanser payu dara dengan ada mungkinnya kehadiran ejen anti-kanser.

ACKNOWLEDGEMENTS

My sincere gratitude goes to my supervising committee chairman, Associate Professor Dr. Radzali Muse, from Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia (UPM), for his patient supports in guides and constructive advices in completing my work. Besides that, I would like to express my appreciation for the chances given by him for me to attend those valuable seminars and conferences during the study.

Great thanks to committee members Associate Professor Dr. Johari Ramli and Dr. Mohd. Yunus Shukor from Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, UPM, Associate Professor Dr. Mohd. Aspollah Sukari from Department of Chemistry, Faculty of Science, UPM. Their valuable suggestions and advices contributed to the completion of my study.

This multidisciplinary study could not be accomplished without the financial support from Intensification of Research in Priority Area (IRPA) programme, provided by the Ministry of Science Technology and Innovation (MOSTI). Not forget to thank Department of Biochemistry for providing the laboratory facilities and Department of Chemistry for providing the facility to carry out GC-MS analysis. At the same time, I would like to show my appreciation to Dr. Muhajir Hamid for providing the laboratory facilities to conduct cytotoxicity test. I am also being grateful for the helps given by the lab assistants and department stuffs.

Special thanks to my lab mates and friends, who provided supports, assistance and encouragement to me along my lab works and study. I would like to show my appreciation to Mr. Tung Sow Hoong for his technical support through out the study and leading me in the completion of this thesis. Last but not least, my deepest thanks to my family members, especially my parents, Lim Choon Huat and Tan Ah Jah, for their continuous supports and understanding.

I certify that an Examination Committee met on 3rd November 2004 to conduct the final examination of Lim Lay Sean on her Master of Science thesis entitled "Analysis of Flavonoids and Essential Oils from *Clausena excavata* and Their Medicinal Properties" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

MUHAJIR HAMID, Ph.D.

Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

MOHD. ARIF SYED, Ph.D.

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

MOHD. PUAD ABDULLAH, Ph.D.

Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

SHAIDA FARIZA SULAIMAN, Ph.D.

Associate Professor School of Biological Sciences Universiti Sains Malaysia (Independent Examiner)

ZAKARIAH ABD. RASHID, Ph.D.

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee are as follows:

RADZALI MUSE, Ph.D.

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

JOHARI RAMLI, Ph.D.

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

MOHD. ASPOLLAH SUKARI, Ph.D.

Associate Professor Faculty of Science Universiti Putra Malaysia (Member)

MOHD. YUNUS SHUKOR, Ph.D.

Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

AINI IDERIS, Ph.D.

Professor/Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

I hereby de	clar	e that the the	sis is based	lon	my o	riginal	work e	exce	ot for o	quot	ations
and citation	s, w	hich have bee	en duly ackı	nowl	edge	d. I also	declar	e tha	at it has	s no	t been
previously	or	concurrently	submitted	for	any	other	degree	at	UPM	or	other
institutions.											

Date:

TABLE OF CONTENTS

			Page
A A A	BSTI BSTI CKN PPR	CATION RACT RAK IOWLEDGEMENTS OVAL ARATION	ii iii v vii ix xi
		OF TABLES	XV
		OF FIGURES	xvi
L	151 (OF ABBREVIATIONS	XIX
C	HAP	TER	
1	INT	TRODUCTION	1
		Introduction	1
	1.2	Objectives of the Studies	9
2	LIT	ERATURE REVIEW	10
	2.1	Plant Secondary Metabolites	10
	2.2	Brief Description about Clausena excavata or 'Cerek Hitam'	11
		2.2.1 Bioactive Constituents from <i>Clausena excavata</i> and Their	
	2.2	Biological Activities	13
	2.3	Flavonoids 2.3.1 Biosynthesis of Flavonoids	16 18
		2.3.2 The Role of Flavonoids in Plants	21
		2.3.3 Flavonoids and Human Health	22
		2.3.4 Solubility Characteristics of Flavonoids	23
		2.3.5 Flavonoid Extraction	24
		2.3.6 Technique of Flavonoid Separation and Identification	25
		2.3.6.1 Paper Chromatography	25
		2.3.6.2 Thin Layer Chromatography2.3.6.3 High Performance Liquid Chromatography	26 27
		2.3.6.4 Ultraviolet-Visible Absorption Spectroscopy	28
	2.4	Volatile Compounds	31
		2.4.1 Biosynthesis of Terpenoids	32
		2.4.2 Essential Oils	34
		2.4.2.1 Biological Activities of Essential Oils	37
		2.4.3 Techniques in Sample Preparation and Identification of	20
		Volatile Compounds 2.4.3.1 Headspace Technique	38 38
		2.4.3.2 Solvent Extraction	40
		2.4.3.3 Simultaneous Distillation Extraction	40
		2.4.3.4 Simultaneous Distillation Adsorption	41
	_	2.4.3.5 Gas Chromatography	41
	2.5	Medicinal Properties	42
		2.5.1 Antioxidant Activity 2.5.1 Free Radical Scavenging Activity of Flavonoids	43 45

		252	2.5.1.2 Methods in the Evaluation of Antioxidant Activity Antibacterial Activity	47 47
		2.3.2	2.5.2.1 Antimicrobial Mechanisms	50
			2.5.2.2 Microbial Susceptibility Test	51
		253	Anticancer Activity	52
		2.3.3	2.5.3.1 Anticancer Mechanisms	56
			2.5.3.2 Human Cancer Cells	57
			2.5.3.3 Cytotoxicity Measurement	57
3			ALS AND METHODS	59
			Materials	59
		Reage		60
			etion of <i>C. excavata</i> Leaves with Different Organic Solvents	60
			mination of Total Phenolics	61
			rsis of UV-Visible Spectrophotometry	61
	3.6		sed Phase-High Performance Liquid Chromatography	62
			Preparation of Standard Flavonoids	62
			Sample Extraction and Hydrolysis Conditions	63
			Apparatus for HPLC Analysis Conditions for Reversed Phase-High Performance	63
		3.0.4	Liquid Chromatography	64
	3 7	Extrac	etion and Identification of Essential Oils	64
	5.1		Fresh Sample Extraction	64
			Apparatus for Gas Chromatography-Mass Spectrometry	65
			Analysis Conditions for Gas Chromatography-Mass	
			Spectrometry	66
	3.8	Deter	mination of Antioxidant Activity	66
			Preparation of Sample and Standards	67
		3.8.2	Ferric Thiocyanate (FTC) Method	67
		3.8.3	Thiobarbituric Acid (TBA) Method	68
			Conjugated Diene (CD) Method	68
			1,1-diphenyl-2-picrylhydrazyl (DPPH) Scavenging	69
	3.9		mination of Antibacterial Activity	70
			Preparation of Mueller Hinton Broth	70
			Preparation of Nutrient Agar	71
			Bacterial Strains and Cultures	71
			Preparation of Antibacterial Agents	72
			Preparation of Paper Discs	72
			Disc Diffusion Method Microdilution Broth Method	72 73
	3 10		mination of Cytotoxic Activity	73 74
	5.10		Cell Lines	74
			Preparation of Medium	74
			Preparation of Samples	75
			Preparation of Tamoxifen (Positive Control)	75
			Preparation of MTT (Tetrazolium Salt) Solution	76
			Cell Culture Technique	76
			Cell Storage	77
			Quantification of Cells	77
		3.10.9	Cytotoxicity Assay	78

4	RES	SULTS	S AND DISCUSSION	79
	4.1	Dried	Yield and Total Phenolics of Leaf Crude Extracts	79
	4.2	Comp	parison of Methods in the Determination of Gallic Acid	
		Conte	ent	82
	4.3	Analy	ysis of Flavonoid Compounds Using UV-Visible	
		Spectr	ophotometry Method and RP-HPLC Technique	83
	4.4	Yield	and Composition of Essential Oils	90
	4.5	Antio	xidant Activity	95
		4.5.1	Ferric Thiocyanate (FTC) Method	95
		4.5.2	Thiobarbituric Acid (TBA) Method	97
		4.5.3	Conjugated Diene Method	98
		4.5.4	1,1-diphenyl-2-picrylhydrazyl (DPPH) Scavenging	101
	4.6	Antib	acterial Activity	106
		4.6.1	Antibacterial Activity of Standard Flavonoids	106
		4.6.2	Antibacterial Activity of Different Crude Extracts of	
			C. excavata Leaf	109
		4.6.3	Antibacterial Activity of Essential Oils Extracted from	
			C. excavata	115
	4.7	Cytot	oxic Activity	119
		4.7.1	Cytotoxicity of Methanol Crude Extract of Leaves and	
			Essential Oils	120
5	CO	NCLU	SION	125
R	EFE!	RENC	ES	129
A	APPENDICES			145
Pl	PUBLICATIONS			
B	BIODATA OF THE AUTHOR			160

LIST OF TABLES

Table		Page
1	A selection of frequently encountered flavonoid glycones, their trivial names and structures.	17
2	Percentage of yield of dried crude extracts of <i>C. excavata</i> leaves extracted with solvents of different polarity.	80
3	Comparison of the methods used in the determination of gallic acid content of different crude extracts.	83
4	Maximal UV absorption and amounts of each flavonoid compound identified in methanol crude extract of <i>C. excavata</i> leaves by UV-visible spectrophotometry method.	84
5	(a) Quantification of each flavonoid compound detected in unhydrolysed and HCl hydrolysed methanol extract of <i>C. excavata</i> leaves by RP-HPLC method at UV detection of (a) 360 nm, and (b) 280 nm.	87
6	Yield of essential oils from different part of <i>C. excavata</i> .	90
7	Retention time, percentage (%) of chemical components identified in the essential oil extracted from (a) fruits of <i>C. excavata</i> , and (b) leaves of <i>C. excavata</i> .	92
8	IC ₅₀ values (mg/ml) for different crude extracts of <i>C. excavata</i> leaves and five selected standard flavonoids.	103
9	Antibacterial activity of four selected standard flavonoids.	108
10	Antibacterial activity of different crude extracts of <i>C. excavata</i> leaves, by determining their (a) diameter of inhibition zone (mm), and (b) minimum inhibitory concentration (MIC)	110
	(μg/ml).	110
11	Antibacterial activity of essential oils extracted from fresh fruits and leaves of <i>C. excavata</i> , by determining their (a) diameter of inhibition zone (mm), and (b) minimum inhibitory concentration (MIC) (µg/ml).	116
12	The selected cancer cells growth inhibitory activity of methanol crude extract of leaf, and essential oils extracted from fruit and leaf of <i>C. excavata</i> .	121

LIST OF FIGURES

Figure		Page
1	A Clausena excavata tree with fruits.	12
2	A generic structure of flavonoid.	16
3	Biosynthesis of flavonoids in plants.	20
4	Ultraviolet-visible absorption spectra of different flavonoid types with equivalent hydroxylation patterns.	30
5	Biosynthesis of terpenoids in plants.	33
6	Some common chemical structures of (a) monoterpene, and (b) sesquiterpene.	35
7	Structural features of flavonoids with a high radical scavenging activity.	46
8	Scavenging of DPPH (free radical) by a flavonoid.	46
9	(a) Healthy leaves, and (b) mature fruits of Clausena excavata.	59
10	Modified Lickens and Nickerson apparatus for simultaneous distillation extraction (SDE).	65
11	Total phenolics (as GAEs) of different solvent extracts of <i>C. excavata</i> leaves.	81
12	Absorbance values of different crude extracts of <i>C. excavata</i> leaves using FTC method.	96
13	Absorbance values (on the sixth day) of different crude extracts of <i>C. excavata</i> leaves using TBA method.	98
14	Antioxidant activity of different crude extracts of C . excavata leaves at two different concentrations of 200 μ g/ml and 1000 μ g/ml after 15 hours incubation times.	100
15	DPPH inhibition percentage (%) of different crude extracts of <i>C. excavata</i> leaves.	102
16	DPPH inhibition percentage (%) of five selected standard flavonoids.	105
17	Antibacterial activity of selected standard flavonoids against <i>Micrococcus luteus</i> (a gram-positive bacterium).	108

18	Antibacterial activity of different crude extracts of <i>C. excavata</i> leaves against <i>Micrococcus luteus</i> , using (a) disc diffusion method, and (b) microdilution broth method.	111
19	Antibacterial activity of essential oils extracted from different parts of <i>C. excavata</i> against <i>Micrococcus luteus</i> , using (a) disc diffusion method, and (b) microdilution broth method.	116
20	Cytotoxic activity of essential oil extracted from <i>C. excavata</i> fruits against MCF-7 cell line (breast cancer cells), using MTT assay.	122
21	A standard graph of gallic acid (μg/ml) determined at wavelength of 765 nm in Follin-Ciocalteu method	145
22	A standard graph of gallic acid (mg/ml) determined at wavelength of 272 nm in UV-visible spectrophotometry method	146
23	A standard graph of apigenin (mg/ml) determined at wavelength of 334 nm in UV-visible spectrophotometry method	147
24	A standard graph of kaempferol (mg/ml) determined at wavelength of 362 nm in UV-visible spectrophotometry method	147
25	A standard graph of quercetin (mg/ml) determined at wavelength of 372 nm in UV-visible spectrophotometry method	148
26	A standard graph of rutin (mg/ml) determined at wavelength of 360 nm in UV-visible spectrophotometry method	148
27	HPLC chromatogram of standard flavonoids, and UV detection at 360 nm.	149
28	HPLC chromatogram of methanol extract of <i>C. excavata</i> leaves before HCl hydrolysis, and UV detection at 360 nm.	150
29	HPLC chromatogram of methanol extract of <i>C. excavata</i> leaves after HCl hydrolysis, and UV detection at 360 nm.	151
30	HPLC chromatogram of standard flavonoids, and UV detection at 280 nm.	152
31	HPLC chromatogram of methanol extract of <i>C. excavata</i> leaves before HCl hydrolysis, and UV detection at 280 nm.	153

32	HPLC chromatogram of methanol extract of <i>C. excavata</i> leaves after HCl hydrolysis, and UV detection at 280 nm.	154
33	Cytotoxic effect of tamoxifen (positive control) on MCF-7 cell line.	155
34	Cytotoxic effect of tamoxifen (positive control) on HeLa cell line.	155
35	Cytotoxic effect of methanol crude extract of <i>C. excavata</i> leaves on MCF-7 cell line.	156
36	Cytotoxic effect of <i>C. excavata</i> fruit oils on MCF-7 cell line.	156
37	Cytotoxic effect of <i>C. excavata</i> leaf oils on MCF-7 cell line.	157
38	Cytotoxic effect of methanol crude extract of <i>C. excavata</i> leaves on HeLa cell line.	157
39	Cytotoxic effect of <i>C. excavata</i> fruit oils on HeLa cell line.	158
40	Cytotoxic effect of <i>C. excavata</i> leaf oils on HeLa cell line.	158

LIST OF ABBREVIATIONS

Abs - absorbance

AIDS - acquired immunodeficiency syndrome

AR - analytical reagent

BHA - butylated hydroxyanisole BHT - butylated hydroxytolune

CO₂ - carbon dioxide DMSO - dimethylsulphoxide

DPPH - 1,1-diphenyl-2-picrylhydrazyl

FBS - foetal bovine serum
FID - flame ionized detector
FTC - ferric thiocyanate

g - gram

GC - gas chromatography HCl - hydrochloric acid

H₂O - water

HPLC - high performance liquid chromatography

IC₅₀ - 50 % inhibitory concentration

i.d. - internal diameter

kg - kilogram

LDL - low density lipoprotein

M - Molar MeOH - methanol mg - milligram

MIC - minimum inhibitory concentration

min - minute

MS - mass spectrometry

MTT - 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl-tetrazolium bromide

NCCLS - National Committee for Clinical Laboratory Standards

nm - nanometer
OD - optical density
ppm - parts per million

PUFAs - polyunsaturated fatty acids ROS - reactive oxygen species

RP - reversed phase

RPMI 1640 - Roswell Park Memorial Institute 1640 SDA - simultaneous distillation adsorption SDE - simultaneous distillation extraction

TBA - thiobarbituric acid
TBQH - tert-butylhydroquinone
TFA - trifluoroacetic acid

TLC - thin-layer chromatography

UV - ultraviolet
μg - microgram
μl - microliter
% - percent

°C - degree Celsius

CHAPTER 1

INTRODUCTION

1.1 Introduction

Two hundred years of modern chemistry and biology have described the role of primary metabolites in basic life functions such as cell division and growth, respiration, storage and reproduction. In biology, the concept of secondary metabolite can be attributed to Kossel (1891). He was the first to define these metabolites as opposed to primary metabolites. Thirty years later, an important step forward was made by Czapek (1921) who dedicated an entire volume of his 'plant biochemistry' series to what he named 'end products'. According to him, these products could well derive from nitrogen metabolism by what he called 'secondary modifications' such as deamination. Compared to the main molecules found in plants, these secondary metabolites were soon defined by their low abundance, often less than 1% of the total carbon, or a storage usually occurring in dedicated cells or organs.

In the middle of the 20th century, improvement of analytical techniques such as chromatography allowed the recovery of more and more of these molecules, and this was the basis for the establishment of the phytochemistry discipline. The improvement of biochemical and biotechnological techniques, and also the rise of molecular biology have been clearly demonstrated that secondary metabolites play a major role in human health and adaptation of plants to their environment.

Plant secondary compounds are usually classified according to their biosynthetic pathways. Three large molecule families are generally considered: (i) phenolics (e.g. flavonoids), (ii) terpenes (e. g. monoterpenes) and steroids, and (iii) alkaloids (Harborne, 1999).

Plant phenolics are an important group of secondary metabolites, which have diverse medicinal applications. Flavonoids are a broad class of low molecular weight secondary plant phenolics characterised by the flavan nucleus. Flavonoids are widely distributed in the leaves, seeds, barks and flowers of plants and over 4,000 flavonoids have been identified. Flavonoids have many functions in the biochemistry, physiology and ecology of plants, and they are important in both human and animal nutrition. In plants, flavonoids have functions in protecting against UV light (UV-B screening pigments), in warding off pathogenic microorganisms (phytoalexins) or pests (antifeedants), in the fertility and germination of pollen, in activating bacterial nodulation genes (nitrogen fixation) and in regulating plant growth and enzyme activity. Plant coloration is not only attractive for pollinators and seed distribution, but also provides aesthetically valuable characteristics for humans. The antioxidant activity of flavonoids towards free radicals and reactive oxygen species, and their potential oestrogenic and anticancer activity, such as antiproliferation, promotion of differentiation and apoptosis, draws attention to their health-protecting role in human diet and animal feed (Harborne and Williams, 2000).

Essential oils are odorous products obtained from natural raw materials such as leaves, fruits, flowers, roots and wood of many seasonal or perennial plants. They are generally of complex composition and contain alcohols, aldehydes, ketones,

phenols, esters, ethers, and terpenes in varying proportions. An estimated 3,000 essential oils are known of which approximately 300 are of commercial importance. On account of their aroma and highly volatile nature, essential oils have been traditionally used as basic raw materials in perfumes and flavouring. They are used in the preparation of beverages, medicines, and personal care and household products such as cosmetics, toiletries and cleaning preparations. They are also used in antiseptics, deodorants, disinfectants and in flavouring of foods and beverages. Essential oils such as monoterpenes are commonly accumulated in *Citrus* species of the Rutaceae family and other families such as Labiatae, Pinaceae and Umbelliferae. Monoterpenes have boiling point at 140-180 °C. They can be separated using steam or hydrodistillation and identified using gas chromatography / mass spectrometry technique.

In the industrialised countries, people are seeking alternative herbal medicine because of the side effect from the strong modern drugs. According to World Health Organisation (WHO), 70 – 90 % of world population especially from developing countries use plant remedies for their health care. It has been indicated that Peninsular Malaysia and the neighbouring islands have more than 6,000 to 7,000 species of higher plants that have therapeutic or medicinal properties. They have been used for many generations in various systems of traditional medicines. Malaysia has over 4,000 Chinese herbal stores which import medicinal plants from Indonesia, China and India, and the locally available species are neglected or underutilised.

Plants are potential sources of natural antioxidants such as e. g. flavonoids and other polyphenols. Natural antioxidants have importance for nutritional and therapeutic applications. Understanding the nutritional and therapeutic role of natural antioxidants is essential for the development of functional foods, which refers to the improvement of conventional foods with added health benefits. This is becoming very significant at a time when food is playing a major role in disease prevention in a global population that is projected to increase to 9 billion by the year 2050. Disease prevention and management through the diet are potentially the most effective tools to improve health and reduce the increasing health-care costs for the expanding global population (Shetty, 2003).

As dietary compounds, flavonoids are widely known as effective antioxidants that inhibit lipid peroxidation and offer protection against oxidative damage to membrane functions. Currently use of synthetic antioxidants has been suspected to cause or promote negative health effects. Both powerful synthetic antioxidants of butylated hydroxyanisole (BHA) and butylated hydroxytolune (BHT) have been widely used for many years to retard lipid oxidation. Nevertheless, these synthetic antioxidants are suspected to be carcinogenic and have been restricted use in foods (Madavi & Salunkhe, 1995). Hence, there is a need to substitute them with naturally occurring antioxidants.

Antioxidants are important not only for food protection but also as a defence mechanism of living cells against oxidative damage. It is well known that humans, as they grow older, become less active, have an increased probability of illness, and generally experience a loss of optimum function of all physiological systems. Lipid peroxidation causes serious damage to the human body. Many researchers have shown that lipid peroxidation *in vivo* is the primary cause of many of the cardiovascular diseases such as atherosclerosis, and also in cancer and aging. The endogenous antioxidants distributed in and around living cells, which regulate the various oxidation-reduction reactions, are seen as a potential class of determinants of longevity (Cutler, 1984). In order to replenish the age-induced loss in the capability of endogenous antioxidant defence mechanisms, there is a need to identify new phytochemicals that could be made readily available by the regular intake of conventional foods.

Recent publications indicate that there is much evidence that plant antioxidants play an important role in biological systems in vitro as agents for antioxidative defence. Antioxidant compounds have already been found in numerous plant materials such as oilseeds, cereal crops, vegetables, fruits, leaves and leaf wax, barks and roots, spices, herbs and crude plant drugs (Ramarathnam *et al*, 1995). Like many green leaves, rosemary contains β-carotene, ascorbic acid, tocopherol, and selenium. Many other antioxidants could complement the conventional vitamins. Classically, rosemary is considered a good antioxidant herb. It contains close to two dozen named antioxidants. Antioxidants from rosemary can be competed with those powerful synthetic antioxidants such as BHA and BHT (Duke and Bogenschutz-Godwin, 1999).

Microbial activity is a primary mode of deterioration of many foods and is often responsible for the loss of quality and safety. Concern over spoilage and poisoning of foods by microorganisms is increasing due to the increase in outbreaks of food