

UNIVERSITI PUTRA MALAYSIA

CHARACTERIZATION AND VIRULENCE STUDIES OF MOTILE AEROMONADS ISOLATED FROM CLARIAS BATRACHUS AND C. GARIEPINUS AND THEIR IMMUNIZATION POTENTIAL

HAMBALI SUPRIYADI

FPSS 1990 1



CHARACTERIZATION AND VIRULENCE STUDIES OF MOTILE AEROMONADS ISOLATED FROM CLARIAS BATRACHUS AND C. GARIEPINUS AND THEIR IMMUNIZATION POTENTIAL

by

Hambali Supriyadi

Thesis Submitted in Partial Fulfillment of the Requirement for the Degree of Master of Science in the Faculty of Fisheries and Marine Science Universiti Pertanian Malaysia

April 1990



ACKNOWLEDGEMENTS

It is my special pleasure to extend my deepest appreciation and gratitude to Associate Professor Dr Mohamed Shariff bin Mohamed Din for his untiring supervision and encouragement to make this project a reality, Dr Hassan Mohd Daud for supervisory help and encouragement and Ms Mariana Nor Shamsuddin for her guidance and help during the project.

I thank Dr Atmadja Hardjamulia, the Director of the Research Institute for Freshwater Fisheries (RIFF), Bogor, Indonesia, for his kind consideration in generously granting the necessary leave and other facilities to enable me to complete this project.

My sincere thanks to International Development Research Centre (IDRC), Canada, for providing me financial support to undertake the course work at Universiti Pertanian Malaysia (UPM) and research project which was conducted in Indonesia, under Fish Parasites Project Phase II.

I am also thankful to Dr Akhmad Rukyani, coordinator, Fish Disease Section, RIFF, Indonesia for the encouragement and support during my in-country research and Messrs Komar, Mariyono and Taukhid for their assistance during my in-country research in Indonesia.

I am indebted to my all friends and colleagues both at RIFF and UPM who helped me in different ways to overcome the difficulties of uncertain periods that I went through during this project.



Last but not least I deeply acknowledge the help, encouragement and patience of my wife Siti Masriyah, my two sons Ivan and Anto and daughter Maylanie without which this would never become a success.



TABLE OF CONTENTS

		Page
ACKNOWLED	Gements	ii
LIST OF T	ABLES	vi
LIST OF F	IGURES	ix
ABSTRACT	• • • • • • • • • • • • • • • • • • • •	xii
ABSTRAK		xv
CHAPTER		
I	GENERAL INTRODUCTION	1
	Background	1
	Objectives	3
II.	REVIEW OF LITERATURE	5
	Taxonomy	5
	Pathogenicity	7
	Transmission	8
	Prevention and Control	10
III	CHARACTERIZATION OF MOTILE AEROMONADS	
	ISOLATED FROM CLARIAS BATRACHUS AND C. GARIEPINUS	14
	Introduction	14
	Materials and Methods	16
	Results	18
	Discussion	24



IV	DETERMINATION OF VIRULENCE OF SELECTED MOTILE AEROMONADS BY PLATE ASSAY	
	TECHNIQUE AND LD ₅₀	29
	Introduction	29
	Materials and Methods	31
	Results	34
	Discussion	44
v	SENSITIVITY OF SOME ANTIBIOTICS TO MOTILE AEROMONAD ISOLATES	48
	Introduction	48
	Materials and Methods	50
	Results	52
	Discussion	56
VI	VACCINATION OF CLARIAS BATRACHUS AGAINST AEROMONAS HYDROPHILA	60
	Introduction	60
	Materials and Methods	63
	Results	65
	Discussion	77
VII	CONCLUSIONS AND SUGGESTIONS FOR FURTHER WORK	82
REFERENCE	S	86
APPENDIX	•••••	96
BIOGRAPHIC	CAL SKETCH	112



LIST OF TABLES

Table		Page
1	Morphological Characteristics of 30 Motile Aeromonad Isolates From Clarias spp	. 19
2	The Effect of Salt Concentrations and Temperatures on the Growth of Motile Aeromonad Isolates	. 21
3	Biochemical Characteristics of 30 Motile Aeromonad Isolates And Type Strain ATCC 7966	. 22
4	Percentage Differences of Biochemical Characteristic of 30 Motile Aeromonad Isolates Compared To The Phenotypic Characteristics Given by Popoff and Lallier (1984)	. 23
5	Percentage Differences of Biochemical Characteristics of 30 Motile Aeromonad Isolates Compared To Typed Strain Aeromonas hydrophila ATCC 7966	. 23
6	Percentage of Positive Enzymatic Activity of 30 Motile Aeromonad Isolate on Different Media	
7	Zone Ratio of Plate Assay Technique of 30 Motile Aeromonad Isolates	. 36
8	Enzymatic Activity of 30 Motile Aeromonad Isolates and the Significant Values	. 37
9	Primary Screening for Pathogenicity of 30 Motile Aeromonad Isolates on C. gariepinus	. 38
10	The Effect of Crude ECP of Motile Aeromonad Isolates on <i>C. gariepinus</i> following Intraperitoneal Injection	. 39



11	The Effect of Different Dose of Crude ECP of Selected Motile Aeromonad Isolates on <i>C. gariepinus</i>	40
12	LD ₅₀ (96 hr) of Selected Motile Aeromonad Isolates on <i>C. batrachus</i> and <i>C. gariepinus</i>	41
13	Crude ECP Protein Content of Four Motile Aeromonad Isolates	41
14	The Inhibition Zone Diameter and MIC Test of 30 Motile Aeromonad Isolates Against Oxytetracycline Hydrochloride	53
15	The Inhibition Zone Diameter and MIC Test of 30 Motile Aeromonad Isolates Against Antibiotic Enrofloxacin	54
16	The Inhibition Zone Diameter and MIC Test of 30 Motile Aeromonad Isolates Against Antibiotic Flumequin	55
17	The Sensitivity Test of 30 Motile Aeromonad Isolates Against Oxytetracycline Hydrochloride	56
18	The Sensitivity Test of 30 Motile Aeromonad Isolates Against Enrofloxacin	56
19	The Sensitivity Test of 30 Motile Aeromonad Isolates Against Flumequin	56
20	Survival (%) of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged at Two-Weeks Post-Vaccination	65
21	Antibody Titers of Vaccinated and Unvaccinated Fish at Two-Weeks Post-Vaccination	67
22	Survival (%) of Vaccinated and Unvaccinated C. batrachus, Challenged	60



23	Unvaccinated Fish at Four-Weeks Post-Vaccination	69
24	Survival (%) of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged at Six-Weeks Post-Vaccination	71
25	Antibody Titers in Vaccinated and Unvaccinated Fish at Six-Weeks Post Vaccination	72
26	Survival (%) of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged at Eight-Weeks Post-Vaccination	73
27	Antibody Titers in Vaccinated and Unvaccinated Fish at Eight-Weeks Post-Vaccination	74
28	Mean Concentration of Ammonia in The Experimental Tanks	75
29	Mean pH in the Experimental Tanks	76
30	Mean Concentration of Dissolved Oxygen in Experimental Tanks	77
31	Motile Aeromonad Isolates and The Health Status of Their Host	96
32	Relationship Between Number of Bacterial Cells per ml and Its Optical Density	97
33	Phenotypic Characteristics of A. hydrophila, A. caviae and A. sobria (Popoff and Lallier, 1984)	97
34	Percentage Survival of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged At Two-Weeks Post-Vaccination	98
35	Analysis of Variance of Vaccinated and Unvaccinated Fish Challenged At Two-Weeks Post-Vaccination	98



36	Percentage Survival of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged At Four-Weeks Post-Vaccination	99
37	Analysis of Variance of Vaccinated and Unvaccinated fish Challenged At Four-Weeks Post-Vaccination	99
38	Percentage Survival of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged At Six-Weeks Post-Vaccination	100
39	Analysis of Variance of Vaccinated and Unvaccinated Fish Challenged At Six-Weeks Post-Vaccination	100
40	Percentage Survival of Vaccinated and Unvaccinated <i>C. batrachus</i> , Challenged At Eight-Weeks Post-Vaccination	101
41	Analysis of Variance of Vaccinated and Unvaccinated fish Challenged At Eight-Weeks Post-Vaccination	101
42	Concentration of Ammonia In The Experimental Tanks	102
43	Analysis of Variance of Ammonia Concentration In The Experimental Tanks	102
44	Concentration of Dissolved Oxygen In The Experimental Tanks	103
45	Analysis of Variance of Dissolved Oxygen In The Experimental Tanks	103
46	pH of Water In The Experimental Tanks	104
47	Calculation of 96 hr LD_{50} of Isolate No.4 on <i>C. gariepinus</i>	105
48	Calculation of 96 hr LD ₅₀	106



49	Calculation of 96 hr LD_{50} of Isolate No.26 on <i>C. gariepinus</i>	107
50	Calculation of 96 hr LD_{50} of Isolate No.27 on <i>C. gariepinus</i>	108
51	Calculation of 96 hr LD_{50} of Isolate No.4 on <i>C. batrachus</i>	109
52	Calculation of 96 hr LD ₅₀	110



LIST OF FIGURES

Figure		Page
1	Dose Response of Crude ECP of Motile Aeromonad Isolates on <i>C. gariepinus</i> Following Intraperitoneal Injection	42
2	Histogram on Dose Response of Crude ECP of Motile Aeromonad Isolates on <i>C. gariepinus</i> Following Intraperitoneal Injection	43
3	A Map of The Province of West Java Indicating The Sample Collection sites	111



Abstract of thesis submitted to the Senate of Universiti Pertanian Malaysia in partial fulfilment of the requirement for the degree of Master of Science.

CHARACTERIZATION AND VIRULENCE STUDIES OF MOTILE Aeromonas ISOLATED FROM Clarias batrachus AND Clarias gariepinus AND THEIR IMMUNIZATION POTENTIAL

By

Hambali Supriyadi

May 1991

Supervisor: Assoc. Professor Dr Mohamed Shariff

Faculty : Fisheries and Marine Science

Morphological, physico-chemical, biochemical and virulence characteristics of 30 motile *Aeromonas* isolates were examined. Selected isolate were investigated for their ability to immunise fish.

Four Aeromonas hydrophila, ten Aeromonas hydrophila-like and sixteen unspecified Aeromonas spp. were isolated from C. batrachus and C. gariepinus. These two fish species were collected from fish farms at Bogor, Depok, Sukabumi and Bekasi, West Java.

It appears that the virulency of motile aeromonads was associated with the type and amount of extracellular substances produced.

Four isolates were selected for the determination of LD_{50} based on enzymatic activity, extracellular products, and preliminary screening of pathogenicity. The LD_{50} studies indicated that all the four Aeromonas spp. isolates tested were virulent to the target fish C. gariepinus ($LD_{50}=10^4$ cells/ml), two isolates were weakly virulent to A. batrachus ($LD_{50}=10^5$ cells/ml) while the other two isolates were avirulent to A. batrachus ($LD_{50}>10^7$ cells/ml).

All thirty isolates of motile aeromonads were screened for sensitivity to three antibiotics: oxytetracycline hydrochloride, enrofloxacin and flumequin. The results reveal that eleven isolates were (36.7%) very sensitive to oxytetracycline hydrochloride, two (6.7%) were fairly sensitive, eight (26.7%) were moderately resistant and nine (30%) were very resistant. None of 30 isolates were resistant to enrofloxacin; 19 isolates (63.3%) were sensitive and 11 (36.7%) were moderately sensitive. Twenty one isolates (70%) were resistant to flumequin, seven (23.3%) were moderately sensitive and two (6.7%) were sensitive.

Three types of vaccine, composed of whole cell bacterin, broth culture and supernatant of A. hydrophila

were tested. Clarias batrachus were immersed in the vaccine for variable periods (5, 10 and 15 minutes), using a single dose. After a standard time the fish were tested serologically for their ability to produce agglutinating antibody. The results showed that there was a significant difference (P<0.05) in antibody response between fish exposed to broth culture vaccine for 15 minutes and unvaccinated fish at two weeks post-vaccination. The highest level of protection (90%) was detected at the fourth week post-vaccination. However, there was no significant difference (P<0.05) between different types of vaccines used and also between different immersion times. The level of protection then decreased after six weeks.

These studies suggest that fish exposed to Aeromonas culture broth provided effective protection over a limited period of about 2 months.

Abstrak tesis yang dikemukakan kepada Senat Universiti Pertanian Malaysia, sebagai memenuhi sebahagian daripada keperluan untuk mendapat Ijazah Master Sains

KAJIAN MENGENAI PENCIRIAN DAN VIRULEN DARIPADA AEROMONADS MOTIL DARI *C. BATRACHUS* DAN *C. GARIEPINUS* SERTA POTENSI IMMUNISASI

oleh

Hambali Supriyadi

Mei 1991

Penyelia : Professor Madya Dr Mohamed Shariff

Fakulti : Perikanan dan Sains Samudera

Suatu kajian mengenai pencirian morfologi, fiziko-kimia dan biokimia dan virulen bagi 30 pencilan bakteria aeromonads motil telah dijalankan. Pencilan terpilih dikaji untuk menentukan kemampuannya untuk mengimunisasikan ikan.

Empat pencilan Aeromonas hydrophila, sebelas berupa A. hydrophila dan enam belas Aeromonas spp. yang belum pasti spesiesnya telah dipencilkan dari ikan keli C. batrachus dan C. gariepinus. Kedua-dua spesies ikan ini diperolehi dari ternakan ikan keli di Bogor, Depok, Sukabumi dan Bekasi, Jawa Barat.

Kajian menunjukkan virulensi aeromonad motil adalah berkait dengan jenis dan jumlah bahan ekstraselular yang



dikeluarkan. Berdasar pada kajian aktiviti enzimatik, bahan ekstraselular dan tapisan awal empat pencilan dipilih untuk kajian LD_{50} . Hasil kajian mengenai LD_{50} dari empat pencilan bakteri aeromonad motil mendapati bahawa keempat-empat pencilan adalah virulen kepada C. gariepinus ($LD_{50}=10^4$ sel/ml); dua pencilan virulen separa lemah ($LD_{50}=10^5$ sel/ml) dan dua lagi pencilan tidak virulen ($LD_{50}>10^7$ sel/ml).

Kesemua tiga puluh pencilan aeromonads motil ditapis untuk kepekaan terhadap tiga jenis antibiotik iaitu oxytetracycline hydrochloride, enrofloxacin dan flumequin telah digunakan dalam kajian sensitiviti kepada 30 pencilan bakteria Aeromonas motil. Hasil kajian mendapati bahawa sebelas pencilan (36.7%) adalah sangat sensitif terhadap oxytetracycline hydrochloride, dua pencilan (6.7%) sensitif, lapan pencilan (26.7%) tahan dan sembilan pencilan (30%) sangat tahan. Tidak ada pencilan yang tahan terhadap enrofloxacin; sembilan belas pencilan (63.3%) adalah sensitif dan sebelas pencilan (36.7%) adalah separa sensitif. Dua puluh satu pencilan (70%) tahan terhadap flumequin, tujuh pencilan (23.3%) separa tahan, dan dua pencilan (6.7%) tahan.



Tiga jenis vaksin terdiri dari bakterin seluruh sel cell bacterin), kultur kaldu, dan supernatan kultur A. hydrophila dikaji. Clarias batrachus telah dimandikan dalam vaksin dengan tiga peringkat waktu yang berbeza iaitu 5, 10 dan 15 minit dengan menggunakan satu dos. Selepas waktu yang ditetapkan, ikan tersebut telah diuji secara serologi untuk menentukan kemampuannya menghasilkan antibodi aglutinan. Hasil kajian mendapati bahawa ada perbezaan yang bererti (P<0.05) di antara ikan yang diberi vaksin kaldu selama 15 minit dan yang tidak pada dua minggu pasca vaksinasi. Paras ketahanan yang tertinggi (90%) didapati pada minggu ke empat setelah ikan diberi vaksin. Namun demikian tidak ada perbezaan yang bererti (P<0.05) antara ketiga-tiga jenis vaksin yang digunakan, dan juga ketiga peringkat masa perendaman. Paras ketahanan didapati menurun pada enam minggu setelah ikan diberi vaksin.

Kajian ini menunjukkan bahawa ikan yang dimandikan dalam bakterin Aeromonas seluruh sel memberikan ketahanan yang berkesan selama dua bulan.

xvii



CHAPTER I

GENERAL INTRODUCTION

Background

Fish is the primary source of animal protein in Indonesia. The majority of fish consumed originate from marine and freshwater capture fisheries. The contribution of fish through aquaculture production accounted for 12.71% of the total production in 1985 (Rahardjo, 1987) but in recent times the availability of marine and freshwater fishes has declined due to overfishing. The only means of increasing fish production is through intensification of aquaculture. For example, intensification of common carp through the use of a running water system in ponds began in 1971 (Supriyadi, 1986). Intensive fish culture in cages has also been operated intensively in reservoirs and lakes.

Catfish is the other commonly cultured freshwater fish species besides common carp. Intensive culture of walking catfish (*Clarias batrachus*) in Indonesia started only recently. In 1985, catfish production was estimated to be 694 tons, which is equivalent to 1.093 million Indonesian Rupiah (US\$ 1 = 1800 Rupiah) (Rahardjo, 1987).



Intensification of catfish culture is practised by means of increasing the stocking rate and food supplement. Fish farmers are encouraged to culture catfish because it is highly profitable.

African catfish (Clarias gariepinus) was recently introduced into Indonesia and is now being cultured extensively. This species is bigger in size as compared with Clarias batrachus. Clarias gariepinus grows faster, is easier to handle and therefore preferred by farmers for culture.

Intensification of fish culture has resulted in increased outbreaks of fish diseases. The first record in Indonesia of bacterial disease in fish culture conditions was in 1980. Great damage to fish production occurred and losses of common carp broodstock was estimated at two million U.S. dollars (Dana, 1987).

The development of catfish culture in recent years has been hampered by the frequent occurrence of bacterial diseases. Aeromonas hydrophila, Pseudomonas fluorescens and Flexibacter columnaris have been isolated from ulcerative lesions of both cultured and wild catfish in Indonesia (Supriyadi, 1988). Aeromonas hydrophila is the bacterium most frequently and consistently isolated from epizootic ulcerative syndrome (EUS)-positive fish (Torres et al., 1990). Hazen et al. (1978) and Snieszko (1974)



also stated that strains of A. hydrophila are widely distributed in the aquatic environment and are also abundant in sewage and waters that are contaminated with sewage.

Our preliminary observations indicate that one month-old catfish are usually more susceptible to bacterial disease. Bacterial infection is usually observed soon after the fingerlings are released into the ponds. Observations also revealed that catfish that are less than one month old are seldom infected by bacterial disease. However, ectoparasites are commonly found at this stage.

Suprivadi (1986) reported that walking catfish is most susceptible to A. hydrophila infection as compared with giant gouramy (Osphronemus gouramy) and common carp (Cyprinus carpio). Since catfish is more susceptible to bacterial disease, many catfish farmers in Indonesia experienced economic losses due to the frequent occurrence of bacterial infection which caused 90 - 100% mortality.

Objectives

Studies on the identification and characterization of Aeromonas spp. have been made by several workers (Shotts and Bullock, 1975; Popoff and Veron, 1976; Popoff and Lallier, 1984).



Aeromonas spp. isolated in Indonesia have been previously reported (Anon, 1980; Sumawidjaja et al., 1981; Supriyadi, 1988). These studies identified the isolates to the genus level only. The aims of the present study were to characterize motile Aeromonas spp. isolated from two species of diseased and healthy catfish, C. batrachus and C. gariepinus, and to evaluate the immunization potential of a selected isolate. The specific objectives of this study were:

- 1. To isolate Aeromonas spp. from health fish.
- To conduct morphologically, physicochemically and biochemically studies on these isolates.
- 3. To determine the virulence of selected Aeromonas spp. by plate assay technique and LD_{50} .
- 4. To evaluate the sensitivity of Aeromonas spp. against selected antibiotics and
- 5. To assess the immunization potential of A. hydrophila vaccine.



CHAPTER II

REVIEW OF LITERATURE

Taxonomy

Aeromonas hydrophila is phenotypically, serologically, and genetically a heterogenous taxon (Newman, 1983). Strains of A. hydrophila have been responsible for massive mortalities of cultured and feral fish populations.

Snieszko (1957) in the 7th edition of Bergey's Manual of Determinative Bacteriology described three species of motile Aeromonads: Aeromonas liquefaciens, A. punctata and A. hydrophila.

Popoff (1984) has grouped the fish-pathogenic aeromonads into two discrete groups, namely the non-motile psychrophilic species and the motile mesophilic species. The genus is, at present, classified in the family Vibrionaceae. However, on the basis of newly acquired evidence resulting from molecular genetic studies Colwell et al. (1986) have proposed that Aeromonas be removed from the Vibrionaceae and placed into a new family Aeromonaceae.



Classification of the species under the genus Aeromonas in the 8th edition of Bergey's Manual of Determinative Bacteriology (Schubert, 1974) differentiated three species within Aeromonads:

Species : Aeromonas hydrophila

Subspecies : - A. hydrophila subsp. hydrophila

- A. hydrophila subsp. anaerogenes

- A. hydrophila subsp. proteolytica

Species : Aeromonas punctata

Subspecies : - A. punctata subsp. punctata

- A. punctata subsp. caviae

Species : Aeromonas salmonicida

Subspecies : - A. salmonicida subsp. salmonicida

- A. salmonicida subsp. achromogenes

- A. salmonicida subsp. masoucida

Popoff (1984) classified four species under the genus Aeromonas :

- 1. Aeromonas hydrophila
- 2. Aeromonas caviae
- 3. Aeromonas sobria
- 4. Aeromonas salmonicida with three subspecies :
 - A. salmonicida subsp. salmonicida
 - A. salmonicida subsp. achromongenes
 - A. salmonicida subsp. masoucida



Pathogenicity

Aeromonas hydrophila has been isolated from a variety of environmental sources, including water, sediments and from diseased and healthy animals. Strains of A. hydrophila have been associated with a wide range of infections affecting cold and warm blooded animals (Davis, et al., 1978).

De Figueiredo and Plumb (1977) examined the virulence of nine A. hydrophila strains isolated from diseased fish and shrimp from freshwater ponds and from channel catfish (Ictalurus punctatus) fingerlings. Significant differences in the ability to kill catfish were observed between strains isolated from water compared to strains from diseased fish. They concluded that the source of the isolate may be related to its ability to produce disease. Lallier et al. (1980) confirmed these observations when they found that strains of A. hydrophila isolated from healthy and diseased fish were more virulent for rainbow trout than an A. sobria strain isolated from healthy fish. Aeromonas hydrophila have been recognized as the causative agent of "red " disease in amphibians and reptiles (Shotts et al., 1972), snails and cow (Popoff and Lallier, 1984) and man (Davis et al., 1978).

Aeromonas hydrophila may posses virulence factors such as proteases, enterotoxin and hemolysins but their

